Synthesis of an Apionucleoside Family and Discovery of a Prodrug with Anti-HIV Activity

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S Supporting Information

[AB](#page-14-0)STRACT: [We report t](#page-14-0)he synthesis of a family of D- and L-furano-Dapionucleosides, their 3′-deoxy, as well as their 2′,3′-dideoxy analogues with thymine and adenine nucleobases. Single carbon homologation of 1,2-Oisopropylidene-D-glycero-tetrafuranos-3-ulose (15) and optimized glycosylation conditions involving microwave irradiation were key to the successful synthesis of the target compounds. While all target nucleosides failed to show significant antiviral activity, we demonstrated that the triphosphate of 2^{\prime} , 3^{\prime} -deoxy-D-apio-Dfuranoadenosine (1) , in contrast to that of its D-apio-L-furanose epimer 2, was readily incorporated into a DNA template by HIV reverse transcriptase to act as a DNA chain terminator. This led us to convert adenine derivative 1 into two phosphoramidate prodrugs. ProTide 9b was found active against HIV-1 and HIV-2 $(EC_{50} = 0.5-1.5 \mu M)$, indicating that the lack of activity of the parent nucleoside, and possibly also other members of the D-apio-D-furanose nucleoside family must be sought in the inefficient cellular conversion to the monophosphate.

■ INTRODUCTION

Although the pharmacological scope of nucleoside analogues is still expanding, they remain most renowned for their utility as antiviral drugs.¹ 2',3'-Dideoxy-β-D-apio-D-furanonucleosides (DddANs, 1, Figure 1) were synthesized in the early 1990s as potential antiv[ir](#page-14-0)al agents, but were found inactive.^{2−4} However, some of us recentl[y d](#page-1-0)iscovered that the 3′-O-phosphonomethylated adenine (A) and thymine (T) analog[ue](#page-14-0)s 7 exhibit promising anti-HIV properties.⁵ Since these phosphonates act as bioisosteres of the phosphorylated species 8, we decided to reinvestigate the biological [ac](#page-14-0)tivity of these ddANs. We envisioned a synthetic approach that would also give access to the known apionucleosides 3, 6,7 their 3′-deoxy counterparts 2^{8-11} and inadvertently also the D-apio-L-furanose epimers 4– 6. Furthermore, we planned t[o e](#page-14-0)xpand the potential of the 2′[,3](#page-14-0)′[-d](#page-14-0)ideoxyapio nucleosides 1 and 4 as antiviral agents by synthesizing their phosphoramidate prodrugs 9, 10 and 11. These would lead to the intracellular release of the parent nucleotides like $8,^{12,13}$ thereby bypassing the often problematic first phosphorylation step in the conversion to the active triphosphate spec[ies](#page-14-0).^{[14](#page-14-0)}

■ RESULTS AND DISCUSSION

Chemistry. Compounds 16 and 27 were considered valuable intermediates to access the envisaged family of Dfuranoapionucleosides (Scheme 1). They were prepared from 1,2-isopropylidene- α -L-threose (14), which was obtained in six steps from L-ascorbic acid.^{15,16} Interestingly, screening of different oxidation methods^{15,17,[18](#page-1-0)} to convert 14 to ketone 15 indicated that TEMPO-BA[IB \(](#page-14-0)[Bis(acetoxy)-iodo]benzene) oxidation, best known fo[r oxid](#page-14-0)ation of primary hydroxyl groups, was the most effective. Conversion of 15 to 16 is feasible by reacting the former with diazomethane to give a spiro-oxirane,¹⁹ which can then be opened with benzylalkoxide to give $16.^{20}$ To avoid the use of diazomethane, we explored several variati[on](#page-14-0)s of the polarity reversal concept to realize the desired car[bo](#page-15-0)n homologation. Reaction with benzyloxymethyl chloride in the presence of samarium iodide did not yield the desired product, while the corresponding Grignard reaction gave 16 in disappointing yields.²¹ Nucleophilic attack of the ketone with lithiated benzyloxymethyltributyltin afforded 16 in acceptable yield, 22 considering t[he](#page-15-0) propensity of compound 15

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to undergo self-condensation to the aldol dimer. 23 The NMR spectra of 16 were in accordance with reported data,²⁰ and C-3 stereochemistry was further confirmed by a tw[o-d](#page-15-0)imensional (2D) ¹H−¹H NOESY experiment. One-pot acid hy[dro](#page-15-0)lysis and acetylation of 16 gave the triacetylated apiose 17 in a 2:1 α/β anomeric ratio.

Jin et al. reported the conversion of 23 to 27 using Barton− McCombie deoxygenation (BMD).¹⁸ Surprisingly, BMD of 16 afforded 18 instead of 27. This led us to reinvestigate the protocol of Jin et al. on compo[und](#page-14-0) 23, prepared from the $\overline{\rm{h}}$ commercially available $\overline{\rm{21.}}^{\rm{24}}$ In our hands $\overline{\rm{BMD}}$ on $\overline{\rm{23}}$ also gave 18. The stereochemistry was confirmed by 2D $\mathrm{^{1}H-^{1}H}$ NOESY experiment and by inde[pen](#page-15-0)dent synthesis of compound 27. The formation of 18 is explained by radical quenching from the least hindered face, i.e., opposite to the isopropylidene comprising face.²⁵ Furthermore, Jin et al. probably synthesized the enantiomer of 18, since they started from D-galactose, which should l[ead](#page-15-0) to 1,2-O-isopropylidene- α -D-threofuranose (i.e., the enantiomer of 14^{15}). Compound 23 was hydrolyzed and acetylated to give the L-furano analogue of triacetylated apiose 24.

Compound 18 was hydrolyzed and acetylated to give 19 in a 4:1 $(\beta:\alpha)$ anomeric ratio. The anomeric configuration was inferred from the anomeric proton coupling constants, i.e., 0 Hz for the β -isomer and 4.4 Hz for the α -isomer. However, this conversion lacked reproducibility, especially on a larger scale. To overcome this problem, the methyl anomer 20 was synthesized in two steps from 18. Since the coupling constant for anomeric hydrogen is close to 0 Hz, 20 is assumed to be the β -isomer.

Long and co-workers found that 1,2-O-isopropylidine-Lfurano-D-apiose 22 equilibrates into a mixture of the D- and Lfuranose form in acidic acetone, which inspired us to use similar conditions for the epimerization of $25.^{19}$ We hypothesized that the absence of the 3-hydroxyl group would eliminate the repulsive dipole interaction with oxyge[n a](#page-14-0)t position 2, while the

Scheme 1. Synthesis of the D- and L-Furano-D-apiose Coupling Partners 17 and 24 and Their 3-Deoxy Analogues 19, 20, and 28^a

a
Reagents and conditions: (a) TEMPO, BAIB, CH₂Cl₂, rt, 3–4 h, 90%; (b) BOMSnBu₃, n-BuLi, THF, −78 °C, 2 h, 68%; (c) (i) 80% aq. AcOH, 80 °C, 8 h; (ii) Ac₂O, DMAP, pyridine, 55 °C, 16 h, 75%; (d) (i) NaH, CS₂, MeI, THF, 0 °C → rt, 1 h; (ii) Et₃B, Bu₃SnH, toluene, rt, 3–4 h, 68%; (e) (i) 80% aq. AcOH, 80 °C, 8 h; (ii) Ac₂O, DMAP, rt, pyridine, 4 h, 57%; (f) (i) p-TsOH (para-toluenesulfonic acid), MeOH, rt, overnight; (ii) Ac₂O, DMAP, pyridine, $0^{\circ}C \rightarrow$ rt, 4 h, 77%; (g) CH₃COOH-H₂O (2:1), rt, 3 days, 83%; (h) Bu₂SnO, toluene, 140 °C, 2 h, TBAB, BnBr, 100 °C, 18 h, 94%; (i) H₂, Pd/C, MeOH, rt, 5 h, 90%; (j) acetone, conc. H₂SO₄, rt, 1.5 h, Na₂CO₃, 45 min, 73% (after 3 cycles); (k) DMF, NaH, 0 °C, 10 min, BnBr, 0 °C → rt, 18 h, 95%; (l) (i) 80% aq. AcOH, 80 °C, 18 h; (ii) pyridine, Ac₂O, rt, 18 h, 79%.

Table 1. Vorbrü ggen Coupling Conditions

		WOR Conditions OAc BnO 19: $R = Ac$ 20: $R = Me$		OAc BnO 29: $B = Thymin-1-yl$ 30: $B = N^6$ -Bz-Adenin-9-yl	
entry	sugar	silylated nucleobase	reagent	conditions	product $(yield)^a$
	19	т	TMSOTf	$1,2-(CH_2),Cl_2$, rt, 4 h	29 (quant)
$\mathfrak{2}$	19	N^6 -BzA	TMSOTf	1,2-(CH ₂) ₂ Cl ₂ , 40 °C, 48 h	30 $(32\%)^b$
3	20	T	TMSOTf	1,2- $(CH_2)_2Cl_2$ or CH ₃ CN, rt, 4 h	32^c
4	20	T	SnCl ₄	$CH3CN$, rt, 4 h	32^c
	20	N^6 -BzA	SnCl ₄	$CH3CN$, rt, 4 h	
6	20	T	$TMSOTf^d$	CH ₃ CN, 150 °C, 5 min microwave	29 + β -anomer $(78%)^e$
	20	N^6 -BzA	$TMSOTf^d$	CH ₃ CN, 150 °C, 5 min microwave	30 (60%)
	a_{r-1} , $1 + 11$, $n-1$, $n-1$	\cdots	\cdot \cdot \cdot \cdot \cdot \cdot \cdot	\sim 1 \sim \sim \sim 1 \sim 1 \sim	\sim \sim \sim \sim

 a Isolated yields. Dash indicates an unresolvable reaction mixture. ${}^b\!{\rm The}\;2'$ -acetyl analogue of 31 was isolated in equal amount. ${}^c\!{\rm Two}\;di$ astereomers observed by TLC and HRMS analysis. ^d 0.2 equiv of TMSOTf. ^e Inseparable 2:1 mixture of 29 and its β-isomer.

steric interaction of the hydroxymethyl group with the 2 oxygen could result in a favorable D-furano isomer ratio. Hence compound 18 was debenzylated and then equilibrated in acetone−conc. H₂SO₄ to isolate the desired compound 26 in 73% yield after 3 equilibrium cycles. Benzylation of 26 gave 27, which upon hydrolysis and acetylation rendered 3-deoxy-Dapio-D-furanose derivative 28 in good yields.

Having the coupling synthons 17, 19, 20, 24 and 28 in hand, we set out different coupling reaction conditions for 19 and 20 with silylated thymine or $N^{\!\delta}$ -benzoyladenine under Vorbrüggen conditions (Table 1).

Whereas the acetate anomer 19 reacted smoothly at room temperature in 4 h with silylated thymine in the presence of TMSOTf to quantitatively give 29, coupling with N^6 benzoyladenine only afforded the desired coupling product 30 in 32% yield by heating the reaction mixture at 40 °C for 48 h.²⁶ This low yield resulted from the formation of an equal amount of an unknown isomer. ¹H NMR of this isomer su[ff](#page-15-0)ered from peak broadening and indicated the presence of minor impurities. Its UV ($\lambda_{\text{max}} = 331.9 \text{ nm}$) and ¹³C-spectrum was characteristic of an $N¹$ -isomer.^{27,28} After treatment with methanolic ammonia for 2 days, a product was formed that was confidently identified as 31 (Figure [2\). Th](#page-15-0)e binding topology of

Figure 2. Byproducts (or their deprotected form) formed during Vorbrüggen coupling.

the adenine base to the sugar was determined by NMR. A correlation between H-1'and C-2 in a 2D $\mathrm{^{1}H-^{13}C}$ HMBC spectrum indicates that the adenine is either bound via N-1 or $\rm \bar{N}$ -3. In a 2D $\rm ^1H-^{1}H$ NOESY spectrum, NOE cross-peaks were detected between the amide proton and several protons of the sugar moiety, most notably H-1′, H-2′ and ortho-protons of the benzoyl group. In addition to this, ortho-protons of the benzoyl moiety also showed NOE interactions with all up (α -face) protons of the furanose ring. These NOE's are improbable if the base is attached via N-3, since in this case the amide group

and the sugar moiety would be positioned para relative to one another and be spatially too far apart.

Coupling reaction between methylglycoside 20 and silylated thymine (entries 3 and 4), using either TMSOTf or $SnCl₄$, resulted in the formation of two main products that gave spots with comparable intensity on TLC. ESI-HRMS analysis allowed identifying these products as the two diastereomers of $32.^{18,29}$ The condensation reaction of methylglycoside 20 with silylated benzoyladenin[e](#page-14-0) in the presence of [an](#page-15-0)hydrous $SnCl₄$ gave an unresolvable reaction mixture.

Vorbrü ggen coupling of the methyl anomer 20 and silylated thymine under microwave irradiation resulted in an inseparable mixture of two isomeric products in a 2:1 ratio, 30 even after removal of the acetyl and benzyl protecting groups. The ¹H NMR spectrum of the minor isomer 33 sho[we](#page-15-0)d a larger splitting of the anomeric hydrogen doublet (3.2 Hz) compared to the major compound (2.0 Hz), indicating a β -oriented pyrimidine moiety. The gHMBC confirmed the C1′-N1 attachment, while 2D NOESY ratified the relative stereochemistry.

Conversely, microwave-assisted coupling between 20 and silylated N^6 -benzoyladenine gave only the desired α -nucleoside 30 in 60% isolated yield. Clearly, the microwave-assisted coupling with the methyl glycoside is the method of choice to prepare the adenine nucleoside.

Reaction of the triacetyl apiose 24 with silylated thymine under classical Vorbrü ggen conditions provided 34 in very good yield (Scheme 2). Microwave conditions were employed to couple 24 with silylated N^6 -benzoyladenine, affording 35 and minor amounts of t[he](#page-3-0) 2′-OTMS analogue 36. The coupling products 29, 30, 34 and 35 were treated with ammonia in methanol to provide the desired deacetylated products 37−40. Debenzylation of L-furano-D-apio thymine nucleosides 37 and 39 to give the 3′-deoxyapionucleoside 5a and apionucleoside 6a was realized by Pd-catalyzed hydrogenation. The same reaction condition on adenosines 38 and 40 was ineffective, as well as the use of cyclohexene and ammonium formate. This led us to use formic acid as hydrogen source to give 5b and 6b. The byproduct 41 was converted to 5b upon treatment with ammonia in methanol.

Using similar protocols 17 and 28 were converted to 2a,b and 3b in acceptable yields (Scheme 3). Compared to the Lseries, this sequence gave low yields for both thymine and adenine analogues, in particular for [th](#page-3-0)e 3′-deoxy analogues.

a Reagents and conditions: (a) appropriate silylated base, 1,2- $(CH₂)₂Cl₂$, TMSOTf, rt, 4 h, 85% for 34; (b) appropriate silylated base, CH₃CN, 0.2 equiv of TMSOTf, microwave (MW) 300 W, 0 \rightarrow 150 °C, 3 min, 150 °C, 5 min, 40% for 35 and 6% for 36; (c) NH3, MeOH, rt, 4−48 h, 75−96%; (d) H2, Pd/C, MeOH, rt, overnight, 86% for 5a from 37 and 71% for 6a from 39; (e) (i) $Pd(OH)_2$, HCOOH– MeOH (1:1 for 5b, 41 from 38/1:9 for 6b from 40), 55 °C, 5–8 h; (ii) $NH₃$, MeOH, rt, 3 h, 80% over two steps for 5b and 6b.

^aReagents and conditions: (a) silylated thymine, $1,2$ - $\left(\text{CH}_2\right)_2\text{Cl}_2$, TMSOTf, rt, 4 h; (b) silylated N^6 -BzA, CH₃CN, 0.2 equiv of TMSOTf, MW 300 W, 0 \rightarrow 150 °C, 3 min, 150 °C, 5 min; (c) 7 N NH3−MeOH, rt, 4−48 h, 46−97% over two steps, 28% for 43 and 11% of its α -anomer; (d) for 42 and 48, H₂, Pd/C, MeOH, rt, 4 h, 86–89%; (e) (i) Pd(OH)₂, HCOOH–MeOH (1:4), 55 °C, 5 h (ii) NH₂, MeOH, rt, 3 h, 89% for 2b and 3b; (f) thiocarbonyl diimidazole, DMF, 80 °C, 90 min, 89% for 46 and 78% for 47; (g) $P(OCH_3)$ ³, 120 $^{\circ}$ C, 6 h, 90%.

Moreover, coupling of 28 with silylated benzoyladenine produced significant amount of α -isomer (11%), possibly due to participation of the 3′-benzyloxymethyl group, with only 28% of desired β -nucleoside. Since the synthesis of 1a,b via 28 involves many linear steps, we envisaged more convenient access to $2'$,3'-dideoxy- β -D-apio-D-furanonucleosides 1a,b involving Corey-Winter olefination³¹ and stereoselective hydrogenation as key steps. During catalytic hydrogenation the synaddition of the hydrogen ato[ms](#page-15-0) to the double bond is anticipated to occur from the face opposite to the nucleobase.³² Thiocarbonylation of 44 and 45 using thiocarbonylimidazole provided precursor compounds 46 and 47 for Corey−Win[ter](#page-15-0) olefination. Unfortunately, the adenine derivative degraded in trimethylphosphite at 120 °C. The thymine derivative gave the

desired product 48 in excellent yield but the hydrogenation reaction resulted in a mixture of diastereomers 1a and 4a that were inseparable by column chromatography. This forced us to follow the classical route via $2a$, b to the target $2'$, $3'$ -dideoxy analogues.

Initially, the benzyl protected nucleosides 37 and 38 were subjected to a standard Barton−McCombie protocol to give the 2′-deoxygenated products 53 and 54 (Scheme 4). Different

Scheme 4. Synthesis of D- and L-Furano-Ddideoxydihydroapionucleosides^a

a Reagents and conditions: (a) TBSCl, imidazole, DMF, rt, overnight, 82−95%; (b) (i) p-tolylchlorothionoformate, DMAP, ACN, 0 °C → rt, 4 h; (ii) Bu₃SnH, AIBN, toluene, reflux, 2–3 h, 70–90% over two steps; (c) H₂/Pd–C, methanol, rt, overnight, 53 to 4a, 63%; (d) TBAF, THF, rt, 3 h, 55 to 4a, 89%; (e) NH4F, MeOH, 50 °C, 2 days, 86−94%.

hydrogen sources were explored for the subsequent Pdcatalyzed debenzylation, but only the thymine compound 53 could be converted to the desired product 4a with curtailed reproducibility. This was attributed to catalyst poisoning by remaining sulfur residues. Hence, we swapped to TBS as a protecting group to give 49−52 from 5a,b and 2a,b in excellent yields. Compounds 49−52 were submitted to BMD after conversion to the corresponding xanthates with p-tolylchlorothionoformate in the presence of DMAP. These xanthates were isolated after a brief work-up and heated in toluene with tributyltin hydride and azobis(isobutyronitrile) to give the 2′,3′ dideoxyapiose nucleosides 55−58. The TBS group of 55 and 56 was removed using TBAF in THF. However, the removal of the tetrabutylammonium salt to get pure adenosine derivative 4b was not satisfactory; hence, we used NH_4F in methanol at 55 °C for 2 days to give 4a,b and 1a,b in excellent isolated yields.³³

To examine the potential of their monophosphate prodrugs as ant[i-H](#page-15-0)IV agents, apio-dideoxynucleosides 1b and 4b were converted to the corresponding triphosphates 12 and 13, following the method of Caton-Williams (Scheme 5).³⁴ The yield of the D-isomer 12 was low and ¹H NMR indicated internal salt formation. Likewise, the 31P NM[R](#page-4-0) [of](#page-15-0) this compound is uncharacteristic of triphosphate salts, as it showed two broad peaks. The addition of 2 equiv of triethylamine disrupted this internal salt leading to the appearance of the

dideoxyapioadenosine Triphosphates $(D-L-ddAATP)$ 12 and 13^a

^aReagents and conditions: (a) 59, 60, n-Bu₃N, anh. DMF, rt, 1.5 h; (b) (i) $1\overline{b}/4\overline{b}$, anh. DMF, rt, 1.5 h; (ii) I_2 , rt, 20 min, H₂O, rt, 1.5 h, 21% for 12 and 48% for 13.

characteristic triphosphate peaks (see the Supporting Information).

Nucleoside monophosphate prodrugs [\(ProTides\), featuring](#page-14-0) [an a](#page-14-0)lanine as the preferred amino acid,^{35–37} were prepared using two different methods (Scheme 6).³⁸ The thymine analogue 11a was prepared by cou[pl](#page-15-0)i[ng](#page-15-0) 4a with the phosphorochloridate 64a, using tert-butylma[gn](#page-15-0)esium chloride as hydroxyl activator. Under similar reaction conditions compound 4b degraded. Hence, all other analogues were coupled with 64a/b using N-methylimidazole (NMI) as a base in a mixture of anhydrous THF and pyridine as solvents. In all

^aReagents and conditions: (a) $\mathrm{CH_2Cl}_2$, TEA, -78 °C, 30 min, rt, 2 h, 87−96%; (b) t-butyl magnesium chloride, THF, rt, overnight, 22% for 11a; (c) NMI, THF, pyridine, rt, 2 days, 15−88%.

cases, the desired compounds were obtained as a mixture of two diasteroisomers resulting from the two possible configurations of the phosphorus stereo center, as confirmed by the presence of two equal height peaks in the ${}^{31}P$ NMR spectrum. DNA Chain Termination Study Using HIV Reverse Transcriptase. A prerequisite for ProTides to show a good biological profile is that the corresponding triphosphates are good substartes for the final target, such as reverse transcriptase (RT) for HIV. Hence we investigated the ability of

triphosphates 12 and 13 to act as a substrate of HIV-RT in a primer-template assay.³⁹ The template has overhanging T residues to test incorporation of the modified A nucleotide. Figure 3 clearly shows [tha](#page-15-0)t both nucleotides 12 and 13 function

Figure 3. Electrophoregram that shows DNA chain termination through incorporation of dideoxydihydro-D-apio-D-furano-adenosine triphosphate (12) and dideoxydihydro-D-apio-L-furano-adenosine triphosphate (13) by HIV RT. The DNA polymeration mixtures containing 125 nM annealed (labeled) primer-template complex were treated with 125, 500, or 1000 μ M of modified triphosphate (12/13) and 0.03 U μ L⁻¹ HIV RT and incubated at 37 °C. Aliquots were taken after 15, 30, and 60 min. In the control reaction, 50 μ M of natural dATP was used. Samples were separated on a 0.4 mm 20% denaturing polyacrylamide gel, and the bands were visualized using phosphorimaging.

as DNA chain terminators. The D-furano analogue 12 is more efficiently incorporated than its 3′-epimer 13, but compared to natural substrate 2′-deoxyadenosine triphosphate (dATP), requires a higher concentration and longer time for complete incorporation. The characteristics of 12 toward HIV RT render the corresponding ProTides as potentially useful HIV inhibitors.

Enzymatic Assay Using Carboxypeptidase Y. The putative mechanism of activation of ProTides^{40−42} (Figure 4) involves an enzymatic cleavage of the ester (step a) mediated

Figure 4. Putative mechanism of bioactivation for monophosphate prodrugs.

by an esterase- or carboxypeptidase-type enzyme followed by spontaneous cyclization with releasing the phenolate anion (step b) and to open the unstable mixed anhydride ring by water (step c) providing the intermediate metabolite (D/L) 67a/b. The cleavage of the phosphorus−nitrogen bond of the latter (step d) requires a phosphoramidase-type enzyme, perhaps related to human HINT-1, to release the monophosphate form $(D-/L-68a/b)$.

In order to investigate this mechanism of bioactivation for ProTides 9a and 11a,b, we performed an enzymatic experiment incubating the compounds with carboxypeptidase Y enzyme in acetone- d_6 and Trizma buffer (pH = 7.6) recording a ³¹P NMR at specific time intervals. The L-furano series displayed pronounced difference in rate of hydrolysis among two diastereomers, a phenomenon that was observed earlier for ProTides of phosphates³⁸ and phosphonates.⁴³ For instance, one of the diastereoisomer of $11a^{31}P$ NMR = 3.3 ppm, Figure 5, panel A) seems to b[e m](#page-15-0)ore slowly conver[ted](#page-15-0) compared to

Figure 5.³¹P NMR stack spectra for bioactivation study of compounds 11a (A) and 11b (B) using carboxypeptidase Y enzyme. The assignment of the resonance signals to the indicated metabolites was done in analogy with previous studies.³⁵

the other. In fact, after 18 h, it is [st](#page-15-0)ill present, even after the addition of an extra portion of enzyme, while the diastereomer at 3.5 ppm appears fully converted after about 10 min. In contrast, compound 11b $(^{31}P$ NMR = 3.2 and 3.4 ppm, Figure 5, panel B) shows a near complete conversion of both diasteroisomers to the metabolite L-67b (31P NMR = \sim 7.0 ppm) through the intermediate L-65b (^{31}P NMR = ~4.5 ppm) after 1 h, although there again exists a clear difference in kinetics.

Within 20 min after addition of the enzyme compound 9a $(^{31}P$ NMR = 3.5 and 3.7 ppm) was completely converted to the intermediate metabolite \overline{D} -65a (³¹P NMR = 4.5 and 4.8 ppm), which was fully converted to compound D-67a (³¹P NMR = \sim 7.1 ppm) within an hour (Figure 6). In this case no pronounced diastereomeric discrimination by carboxypeptidase

Figure $6.$ ³¹P NMR stack spectra for bioactivation study of compound 9a using carboxypeptidase Y enzyme. The assignment of the resonance signals to metabolites D-65a and D-67a is based on LC−MS experiments.

enzyme was observed. Following the trend for adenine analogue 11b, we assume that 9b would be processed at the least with the rate of thymine analogue 9a.

From this study it is evident that both D- and Lfuranonucleoside ProTides are readily converted to the intermediate metabolite 67.

Biological Evaluation. The 2',3'-dideoxy analogues 1a,b and the $3'$ -deoxy- β -D-apio-D-furanonucleosides 2a,b failed to show both activity against HIV-1,2 and cytotoxicity. Likewise, the $2^{\prime},3^{\prime}$ -dideoxy analogues 4a,b and the 3'-deoxy- β -D-apio-Lfuranonucleosides 5a,b lacked significant activity against HIV-1 and HIV-2 and a panel of other DNA and RNA viruses, and were also devoid of cytotoxicity. The thymine-based ProTides 9a and 10a were also devoid of anti-HIV activity, which might be due to inefficient conversion of the alaninyl d-ddATMP to the corresponding monophosphate by HINT-1-type phosphoramidase enzyme or further kinase mediated conversion to the corresponding triphosphate. Alternatively, the latter may be inefficiently incorporated by HIV RT (Table 2).

Table 2. Antiviral Activity and Cytotoxicity of ProTides 9a,b and 10a,b

	EC_{50} in MT-4 cells (μM)			EC_{50} in CEM cells (μM)		
	$HIV-1$ (NLA.3)	$HIV-2$ (ROD)	CC ₅₀	$HIV-1$ (IIIb)	$HIV-2$ (ROD)	
9a	>250	>250	196	\mathcal{I}		
10a	>250	>250	>250			
9b	0.5	1.0	93	0.5	1.5	
10b	26	24	>250	7.5	38	
R-PMPA	1.7	1.0	>250	3.0	2.5	
"Dashes indicate not performed.						

Interestingly, the 2′,3′-dideoxy-D-apio-D-furanoadenosine phosphoramidate ProTides 9b and 10b combine potent and moderate anti-HIV activity with reasonable selectivity. The benzylester 9b exhibits comparable or even somewhat superior anti-HIV activity to the acyclic nucleoside phosphonate R-PMPA (tenofovir).⁴⁴ The ProTides 9a,b, 10a,b and 11a,b are weak to moderate inhibitors of murine leukemia (L1210), human T-lympho[cyt](#page-15-0)e (CEM) and human cervix carcinoma (HeLa) cell proliferation (Table 3).

Table 3. Cytotoxicity Data of ProTides 9a,b and $10a,b^a$

	L ₁₂₁₀	CEM	HeLa
9a	$113 + 21$	$108 + 11$	159 ± 32
9b	$110 + 17$	$80 + 4$	53 ± 11
10a	>2.50	>2.50	>2.50
10b	$226 + 35$	$204 + 3$	>250
11a	$167 + 85$	113 ± 3	177 ± 103
11b	$79 + 4$	$73 + 5$	$173 + 58$

 ${}^{a}IC_{50}$ in μ M, murine leukemia cells (L1210/0), human T-lymphocyte cells (CEM/0), and human cervix carcinoma cells (HeLa)

■ CONCLUSION

In this study we report the synthesis of a family of Dapionucleosides comprising the A and T members of both possible 3'-epimers of $β$ -D-apiofuranose nucleosides, as well as their 3′-deoxy and 2′,3′-dideoxy analogues. Clues in the synthesis of the desired apionucleosides were a carbon homologation of 1,2-O-isopropylidene-D-glycero-tetrafuranos-3-ulose (15) and optimized glycosylation conditions involving microwave irradiation. In the course of this work, we rectified some anomalies in the structure assignments reported by others.

In accordance with earlier reports the target D-apio-Dfuranose nucleosides failed to show antiviral activity and so did their D-apio-L-furanose epimers. However, the triphosphate of $2^{\prime},3^{\prime}$ -dideoxy- β -D-apio-D-furanoadenosine (12) (in contrast to its D-apio-L-furanose epimer 13) was readily accepted by viral DNA polymerase to act as a DNA chain terminator. This led us to convert the parent A and T nucleosides 1a and 1b into phosphoramidate prodrugs 9 and 10. The A analogues 9b and 10b indeed showed a considerable anti-HIV activity. This indicates that the lack of activity of the parent $2'$, 3'-dideoxy- β -D-apio-D-furanose nucleoside must be the result of inefficient conversion to the monophosphate in the biological assay. This study demonstrates that the large pool of nucleoside analogues that were previously found to lack antiviral activity may contain valuable candidates to be turned into ProTide derivatives exhibiting promising antiviral activity, by efficiently bypassing the first phosphorylation step that is often rate-limiting the intracellular conversion of nucleoside analogues to their bioactive triphosphate derivatives.

EXPERIMENTAL SECTION

Synthesis. All reagents were from standard commercial sources and of analytic grade. Dry solvents were obtained directly from commercial sources and stored on molecular sieves. All reactions were carried out under argon atmosphere using anhydrous solvents unless specified otherwise. Room temperature or rt refers to 25 ± 5 °C. Silica-gel precoated with F254 plates were used for TLC. The spots were examined under ultraviolet light at 254 nm and further visualized by sulfuric acid-anisaldehyde spray. Column chromatography was performed on silica gel (40–63 μ m, 60 Å) or on flash chromatography system. NMR spectra were recorded on a 300 MHz, 500 or 700 MHz spectrometer. Chemical shifts are given in ppm (δ) , calibrated to the residual solvent signals or TMS. Exact mass measurements were performed on mass spectrometer equipped with a standard electrospray ionization (ESI) and modular LockSpray TM interface. Samples were infused in a CH_3CN/H_2O (1:1v/v) mixture at 10 mL/min. The microwave reactions were carried out in Milestone MicroSYNTH Advanced Microwave Synthesis Labstation, equipped with 2×800 W magnetrons (effective maximum output 1500W pulsed/continuous), an optical fiber temperature sensor, a pressure sensor, in continues power mode in a closed PTFE vessel. NMR signals of sugar protons and carbons are indicated with a prime, and signals of base protons and carbons are given without a prime. A combination of gCOSY, gHSQC and gHMBC was used to assign $^1\mathrm{H}$ and $^{13}\mathrm{C}$ peaks, Noesy was used for selected compounds to assign peaks and/or to confirm relative configuration.

3-Oxo-1,2-O-isopropylidene- α -D-erythrofuranose (15).¹⁸ To a solution of compound 14 (1.0 g, 6.24 mmol) in CH_2Cl_2 (12.5 mL) was added bis-acetoxyiodobenzene (BAIB, 2.41 g, 7.5 mmol[\) fo](#page-14-0)llowed by (2,2,6,6-tetramethylpiperidin-1-yl)oxyl (TEMPO, 195 mg, 1.25 mmol) at room temperature under an argon atmosphere. The mixture was stirred at room temperature for 4 h. The contents of the reaction was directly loaded on silica-gel and eluted with 30% EtOAc-hexanes to afford pure product 15 (890 mg, 90%) as a white solid: ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$ δ ppm 1.35 (s, 3H, CH₃), 1.44 (s, 3H, CH₃), 4.03 (dd, $J = 4.1$, 17.6 Hz, 1H, 4-H), 4.29 (s, 1H, 2-H), 4.32 (dd, $J = 0.6$, 17.6 Hz, 1H, 4-H), 6.02 (d, $J = 4.4$ Hz, 1H, 1-H).

1,2-O-Isopropylidene-5-(O-benzyl)-α-D-apio-D-furanose (16).²⁰ To a stirring solution of benzyloxymethyltributlytin (BOMSnBu₃, 5.93 g, 14.4 mmol) in THF (35 mL) at −78 °C under inert conditi[on,](#page-15-0) was added dropwise n-butyllithium (1.6 M in hexanes,19.5 mL, 31.3 mmol) and stirred for additional 1 h. To this mixture was then added dropwise a solution of compound 15 (1.9 g, 12.02 mmol) in 10 mL of THF and stirred at −78 °C for 3 h. The reaction was quenched with saturated NH4Cl solution and by vigorous stirring. EtOAc (100 mL) was then added to facilitate the layer separation. Organic layer was separated, and the aqueous layer was extracted twice with EtOAc (50 mL). Combined organic layers were dried over anhydrous MgSO₄ and evaporated under reduced pressure. The residue was purified by column chromatography eluting with 17% EtOAc-hexanes to afford 16 (2.3 g, 68%) as a white solid: ¹H NMR (300 MHz, CDCl₃) δ ppm 1.37 (s, 3H, CH₃b), 1.58 (s, 3H, CH₃a), 2.85 (s, 1H, 3-OHa), 3.46 (d, $J = 10.3$ Hz, 1H, 5-H), 3.56 (d, $J = 10.3$ Hz, 1H, 5-H), 3.71 (d, $J = 9.1$ Hz, 1H, 4-Ha), 3.80 (d, J = 9.1 Hz, 1H, 4-Hb), 4.39 (d, J = 3.8 Hz, 1H, 2-Hb), 4.54−4.71 (m, 2H, CH2Ph), 5.76 (d, J = 4.1 Hz, 1H, 1-Hb), 7.27−7.40 (m, 5H, CH_2Ph).

1,2,3-Tri-(O-acetyl)-5-(O-benzyl)- α/β -D-apio-D-furanose (17).⁴⁵ A solution of 16 (2.5 g, 8.92 mmol) in 80% aq. acetic acid (25 mL) was stirred at 80 °C for 8 h. The reaction mixture was evaporated t[o gi](#page-15-0)ve the crude intermediate as syrup. This syrup was dissolved in pyridine (20 mL) and DMAP was added (100 mg) followed by acetic anhydride (10 mL, 106 mmol). The solution was stirred at 55 °C for 16 h. Then, the solvent was removed under a vacuum, and the resulting residue was partitioned between EtOAc and water. Organic layer separated, combined organic layer was washed with brine, dried over sodium sulfate, and evaporated. The residue was purified by silicagel column chromatography (15−20% EtOAc-hexanes) to yield 17 (2.45 g, 75%) as a colorless oil as a mixture of $\alpha + \beta$ isomers (2:1): ¹H NMR (300 MHz, CDCl₃) δ ppm 1.96 (s, 3H, major), 2.08 (s, 3H, major), 2.08 (s, 2H, minor), 2.09 (s, 1H, minor), 2.10 (s, 3H, major), 3.75 (d, J = 9.7 Hz, 0.47H, minor), 3.89 (d, J = 10.5 Hz, 1H, major), 3.96 (d, J = 9.7 Hz, 0.5H, minor), 4.05 (d, J = 10.5 Hz, 1H, major), 4.22 (d, $J = 10.3$ Hz, 1H, major), 4.26 (d, $J = 10.5$ Hz, 0.52H, minor), 4.32 (d, $J = 10.5$ Hz, 0.5H, minor), 4.34 (d, $J = 10.3$ Hz, 1H, major), 4.51−4.62 (m, 3H, major and minor), 5.42 (d, J = 4.7 Hz, 0.44H, minor), 5.49 (d, J = 1.2 Hz, 1H, major), 6.08 (d, J = 1.2 Hz, 1H, major), 6.33 (d, J = 4.7 Hz, 0.43H, minor), 7.27−7.41 (m, 7H, major and minor); HRMS (ESI-TOF) m/z [M + Na]⁺ Calcd for $C_{18}H_{22}O_8$ Na 389.1212, found 389.1242.

1,2-O-Isopropylidene-3-deoxy-5-(O-benzyl)-β-D-apio-L-furanose (18).¹⁸ To a solution of 16 (3.5 g, 12.5 mmol) in dry THF (75 mL) was added NaH (60% in mineral oil, 1.5 g, 37.45 mmol) at 0 °C, and the [rea](#page-14-0)ction mixture was stirred at room temperature for 1 h. To this mixture were slowly added CS_2 (11.2 mL, 188 mmol) and MeI (24.0) mL, 375 mmol) and stirred at room temperature for 1 h. The reaction mixture was evaporated to give crude xanthate. The xanthate was suspended in dry toluene (75 mL), triethylborane (19.0 mL, 19.0 mmol, 1.0 M solution in THF) and $n-Bu_3SnH$ (5 mL, 19.0 mmol) were added at room temperature, and the mixture was stirred for further 3 h. The reaction mixture was quenched with water, extracted with EtOAc, dried over anhydrous MgSO₄, filtered and evaporated. The residue was purified by silica gel column chromatography (10%

EtOAc-hexanes) to give 18 (2.26 g, 68%) as a colorless oil: $^1\rm H$ NMR (300 MHz, CDCl₃) δ ppm 1.32 (s, 3H, CH₃a), 1.49 (s, 3H, CH₃b), 2.37−2.52 (m, 1H, 3-Ha), 3.52 (dd, J = 9.2, 7.5 Hz, 1H, 5-H), 3.69 $(dd, J = 11.3, 8.6 Hz, 1H, 4-Hb), 3.78 (dd, J = 9.4, 6.7 Hz, 1H, 5-H),$ 4.01 (dd, J = 8.4, 7.2 Hz, 1H, 4-Ha), 4.46–4.60 (m, 2H, CH₂Ph), 4.65 $(t, J = 4.1 \text{ Hz}, 1H, 2-Ha), 5.83$ $(d, J = 3.8 \text{ Hz}, 1H, 1-Ha), 7.24-7.42$ $(m, 5H, CH₂Ph).$

1,2-Di-O-acetyl-3-deoxy-5-(O-benzyl)-α/β-D-apio-L-furanose (19). A solution of 18 (750 mg, 2.84 mmol) in 80% aq. acetic acid (10 mL) was stirred at 80 °C for 8 h. The reaction mixture was evaporated to give the crude intermediate as a syrup. This syrup was dissolved in pyridine (15 mL) and treated with DMAP (50 mg) and acetic anhydride (2.0 mL, 21.2 mmol). The solution was stirred at room temperature for 4 h. The solvent was removed under a vacuum, and the resulting residue was purified by silica-gel column chromatography (20% EtOAc-hexanes) to yield 19 (500 mg, 57%) as a colorless oil $(\alpha$: β anomeric ratio 1:4). Major isomer: ¹H NMR (300 MHz, CDCl₃) δ ppm 1.94 (s, 3H, Ac), 1.97 (s, 3H, Ac), 2.83−2.96 (m, 1H, 3-H), 3.40 (dd, $J = 9.1, 7.3$ Hz, 1H, 5-H), 3.55 (dd, $J = 9.1, 7.6$ Hz, 1H, 5-H), 3.77 (t, J = 8.8 Hz, 1H, 4-H), 4.17 (t, J = 8.4 Hz, 1H, 4-H), 4.35–4.48 $(m, 2H, CH_2Ph), 5.20$ (d, J = 5.0 Hz, 1H, 2-H), 6.02 (s, 1H, 1-H), 7.18−7.32 (m, 5H, CH₂Ph); ¹³C NMR (75 MHz, CDCl₃) δ ppm 20.5 (CH_3CO) , 21.0 (CH₃CO), 40.1 (3-C), 66.1 (5-C), 70.9 (4-C), 73.3 (CH₂Ph), 76.1 (2-C), 99.7 (1-C), 127.6 (C_o Ph), 127.7 (C_p Ph), 128.4 $(C_m Ph)$, 137.8 $(C_{ipso} Ph)$, 169.4 (CH_3CO) , 169.7 (CH_3CO) ; HRMS (ESI-TOF) m/z [M + Na]⁺ Calcd for C₁₆H₂₀O₆Na 331.1158, found 331.1152.

1-O-Methyl-2-O-acetyl-3-deoxy-5-(O-benzyl)-β-D-apio-L-furanose
(20).¹⁸ A solution of 18 (2.26 g, 8.55 mmol) and p-TsOH (700 mg, 4.06 mmol) in MeOH (60 mL) was stirred at room temperature for 16 h, n[eut](#page-14-0)ralized with TEA and evaporated. The residue was partitioned between EtOAc and water, organic layer separated, dried over anhydrous $MgSO_4$ and evaporated. The residue was purified by column chromatography (20−40% EtOAc-hexanes). The intermediate was dissolved in pyridine (15 mL), acetic anhydride (2.4 mL, 25.2 mmol) and DMAP (200 mg, 1.68 mmol) were added at 0 °C, and the reaction mixture was stirred at room temperature for 4 h. The reaction mixture was evaporated, and partitioned between EtOAc and 10% aq. KHSO4. The organic layer was dried over anhydrous MgSO4, filtered and evaporated. The residue was purified by silica gel column chromatography (15% EtOAc-hexanes) to give 20 (1.85 g, 77%) as a colorless oil: ¹H NMR (300 MHz, CDCl₃) δ ppm 2.00 (s, 3H, 2-OAc), 2.88−3.02 (m, 1H, 3-H), 3.34 (s, 3H, 1-OMe), 3.46 (dd, J = 9.1, 7.3 Hz, 1H, 5-H), 3.62 (dd, J = 9.2, 7.2 Hz, 1H, 5-H), 3.78 (t, J = 8.6 Hz, 1H, 4-H), 4.14 (t, $J = 8.5$ Hz, 1H, 4-H), 4.46 (d, $J = 12.0$ Hz, 1H, CH₂Ph), 4.52 (d, J = 12.0 Hz, 1H, CH₂Ph), 4.83 (s, 1H, 1-H), 5.16 (d, J = 5.3 Hz, 1H, 2-H), 7.27–7.39 (m, 5H, CH₂Ph).

1,2-O-Isopropylidene-β-D-apio-L-furanose (22).¹⁸ Compound 21 (5.0 g, 21.72 mmol) was dissolved in 50 mL of 2:1 acetic acid−water mixture and stirred at room temperature for 3 [days](#page-14-0). Solvents were evaporated in vacuo and silica gel column chromatography of the residue (50% EtOAc-hexanes) afforded the title compound 22 as a white solid (3.4 g, 83%): ¹H NMR (300 MHz, CDCl₃) δ ppm 1.33 (s, 3H, $C(CH_3)_2$, 1.51 (s, 3H, $C(CH_3)_2$), 2.12 (t, J = 5.9 Hz, 1H, 5-OH), 2.69 (s, 1H, 3-OH), 3.71 (dd, J = 6.3, 11.16 Hz, 1H, 4-H), 3.80 (d, J = 9.8 Hz, 1H, 5-H), 3.94 (d, J = 9.4 Hz, 1H, 5-H), 3.96 (dd, J = 5.4, 7.5 Hz, 1H, 4-H), 4.38 (d, J = 3.8 Hz, 1H, 2-H), 5.99 (d, J = 3.7 Hz, 1H, 1-H).

1,2-O-Isopropylidene-5-(O-benzyl)-β-D-apio-L-furanose (23).^{18,20} Compound 22 (3.1 g, 16.3 mmol) and dibutyltin oxide (6.7 g, 26.9 mmol) was dissolved in toluene (120 mL) refluxed at 140 °C for [2](#page-14-0) [h.](#page-15-0) The reaction mixture was allowed to attain 100 °C then added tetrabutylammonium bromide (2.63 g, 8.15 mmol) and benzyl bromide (3.0 mL, 25.26 mmol). The reaction mixture was stirred at this temperature for 18 h. Solvent was evaporated under reduced pressure, and the residue purified by silica gel column chromatography (30% EtOAc-hexanes) to afford 23 (4.3 g, 94%) as a white solid: ¹H NMR (300 MHz, CDCl₃) δ ppm 1.32 (s, 3H, C(CH₃)₂a), 1.48 (s, 3H, $C(CH_3)_2b$), 2.76 (d, J = 0.9 Hz, 1H, 3-OHa), 3.54 (d, J = 9.7 Hz, 1H, 5-H), 3.80 (d, J = 9.7 Hz, 1H, 5-H), 3.82 (dd, J = 9.4, 0.9 Hz, 1H, 4Ha), 3.88 (dd, J = 9.4 Hz, 1H, 4-Hb), 4.35 (dd, J = 3.5, 0.9 Hz, 1H, 2- Ha), 4.57 (d, $J = 12.0$ Hz, 1H, PhCH₂), 4.64 (d, $J = 12.0$ Hz, 1H, PhCH₂), 5.98 (d, J = 3.5 Hz, 1H, 1-Ha), 7.27–7.42 (m, 5H, PhCH₂).

1,2,3-Tri-(O-acetyl)-5-(O-benzyl)-β-D-apio-L-furanose (24). Following the procedure described for the synthesis of 17, (2.5 g, 8.92 mmol) of 23 rendered pure product 24 (2.45 g, 75%) as an oil: ¹H NMR (300 MHz, CDCl₃) δ ppm 1.96 (s, 3H, COCH₃), 2.07 (s, 3H, COCH₃), 2.10 (s, 3H, COCH₃), 3.89 (d, J = 10.5 Hz, 1H, 4-H), 4.05 $(d, J = 10.3 \text{ Hz}, 1H, 4-H)$, 4.22 $(d, J = 10.3 \text{ Hz}, 1H, 5-H)$, 4.34 $(d, J =$ 10.3 Hz, 1H, 5-H), 4.50–4.62 (m, 2H, PhCH₂), 5.49 (d, J = 1.2 Hz, 1H, 2-H), 6.08 (d, J = 1.2 Hz, 1H, 1-H), 7.26–7.40 (m, 5H, PhCH₂).

1,2-O-Isopropylidene-3-deoxy-β-D-apio-L-furanose (25).¹⁸ Compound 18 (3.7 g, 14 mmol) was dissolved in methanol (100 mL), to this was added Pd–C (3.7 g, 10% Pd, wet ~50% H₂O). [Str](#page-14-0)eam of hydrogen gas was bubbled through the reaction mixture for 5 h at room temperature. The catalyst was filtered off, and the filtrate concentrated to give crude product which on purification by silica-gel column chromatography (40% EtOAc-hexanes) rendered 25 (2.2 g, 90%) as a colorless oil: ¹H NMR (300 MHz, CDCl₃) δ ppm 1.34 (d, J $= 0.6$ Hz, 3H, C(CH₃)₂), 1.53 (s, 3H, C(CH₃)₂), 2.21 (br.s, 1H, 5-OH), 2.34 (ddtd, J = 11.4, 6.9, 5.97, 6.0, 4.8 Hz, 1H, 3-H), 3.82−3.91 $(m, 3H, 4-H \text{ and } 5-H's), 3.97 \text{ (dd, } J = 8.5, 7.3 \text{ Hz, } 1H, 4-H), 4.73 \text{ (t, } J$ $= 4.4$ Hz, 1H, 2-H), 5.86 (d, J = 3.8 Hz, 1H, 1-H).

1,2-O-Isopropylidene-3-deoxy-α-D-apio-D-furanose (26). To a solution of compound 25 (2.2 g, 12.63 mmol) in 400 mL of acetone was added concentrated sulfuric acid (2.2 mL), and the mixture was stirred at room temperature for 1.5 h. Then sodium carbonate (14 g) was added and stirred at room temperature for 45 min. Inorganic salts were removed by filtration, and the filtrate concentrated under reduced pressure to afford oil. TLC indicated the conversion in favor of required isomer (roughly 2:1). The title compound is slightly more polar than the starting material (R_f after two runs: 0.35 for 26 and 0.4 for 25; eluent, 2.5% MeOH in CH_2Cl_2). Silica-gel flash column chromatography (0.5−1.5% MeOH in CH_2Cl_2) afforded title compound and starting material. After three cycles 1.6 g (73%) of **26** was procured as a colorless oil: ¹H NMR (300 MHz, CDCl₃) δ ppm 1.31 (d, J = 0.6 Hz, 3H, C(CH₃)b), 1.50 (s, 3H, C(CH₃)a), 1.89 $(br.s, 1H, 5-OH)$, 2.36–2.46 (m, 1H, 3-H), 3.58 (dd, J = 6.6, 3.4 Hz, 2H, 5–CH₂), 3.83 (d, J = 9.1 Hz, 1H, 4-Hb), 4.10 (dd, J = 8.9, 5.1 Hz, 1H, 4-Ha), 4.60 (d, $J = 3.5$ Hz, 1H, 2-H), 5.81 (d, $J = 3.8$ Hz, 1H, 1-H); ¹³C NMR (75 MHz, CDCl₃) δ ppm 26.2 (C(CH₃)b), 26.8 $(C(CH₃)a)$, 48.1 (3-C), 62.0 (5-C), 68.7 (4-C), 82.3 (2-C), 105.6 (1-C), 111.3 $(C(CH_3)_2)$.

1,2-O-Isopropylidene-3-deoxy-5-(O-benzyl)-α-D-apio-D-furanose (27). To an ice cold solution of 26 (1.6 g, 9.2 mmol) in DMF (30 mL) was added NaH (60% in mineral oil, 0.55g, 13.8 mmol) and then benzyl bromide (1.64 mL, 13.8 mmol) dropwise. The reaction mixture was stirred at room temperature overnight. Methanol (5 mL) was added and stirred for further 30 min. The volatile materials were removed under vacuo, and the residue was partitioned between ethyl acetate and water. The organic layer was separated, dried over anhydrous $Na₂SO₄$, evaporated, and the residue purified by column chromatography (5−15% EtOAc in hexanes) to afford 27 (2.3 g, 95%) as a colorless oil: ¹H NMR (300 MHz, CDCl₃) δ ppm 1.31 (d, J = 0.6 Hz, 3H, C(CH₃)b), 1.51 (s, 3H, C(CH₃)a), 2.56 (td, J = 7.5, 5.1 Hz, 1H, 3-Ha), 3.37 (d, J = 7.6 Hz, 2H, 5−CH2), 3.83 (d, J = 8.8 Hz, 1H, 4-Hb) 4.09 (dd, J = 8.9, 5.1 Hz, 1H, 4-Ha) 4.51 (d, J = 3.2 Hz, 2H, PhCH₂), 4.56 (d, J = 3.5 Hz, 1H, 2-Hb), 5.79 (d, J = 3.5 Hz, 1H, 1-Hb), 7.27−7.41 (m, 5H, PhCH₂); ¹³C NMR (75 MHz, CDCl₃) δ ppm 26.3 (C(CH₃)b), 26.9 (C(CH₃)a), 46.1 (3-C), 68.8 (4&5-C), 73.3 (PhCH₂), 82.4 (2-C), 105.6 (1-C), 111.2 ($C(CH_3)_2$) 127.8, 127.9, 128.6, 138.1 (PhCH₂). MS (ESI-TOF) m/z [M + K]⁺ Calcd for $C_{15}H_{20}O_4K$ 303.0999, found 303.1078.

1,2-Di-O-acetyl-3-deoxy-5-(O-benzyl)-α/β-D-apio-D-furanose (28). Following the procedure described for the synthesis of 19, compound 27 (1.3 g, 4.92 mmol) rendered 28 (1.2 g, 79%) as a colorless oil. Mixture of $\alpha+\beta$ (2:1): ¹H NMR (300 MHz, CDCl₃) δ ppm 2.00 (s, major, $C(CH_3)_2$) 2.04 (s, minor, $C(CH_3)_2$) 2.07 (s, minor, $C(CH_3)_2)$ 2.08 (s, major, $C(CH_3)_2)$, 2.56–2.69 (m, major, 3-H) 2.69−2.83 (m, minor, 3-H) 3.46−3.74 (m, major and minor, 5-H) 3.80−3.94 (m, major and minor, 4-H) 4.20−4.34 (m, major and minor, 4-H) 4.51 (s, minor, PhCH₂), 4.54 (s, major, PhCH₂), 5.05 (t, J $= 4.1$ Hz, minor, 2-H), 5.08 (d, J = 2.6 Hz, major, 2-H), 6.13 (s, major, 1-H), 6.33 (d, J = 4.4 Hz, minor, 1-H), 7.27−7.40 (m, major and minor, PhCH₂); HRMS (ESI-TOF) m/z [M + K]⁺ Calcd for $C_{16}H_{20}O_6K$ 347.0897, found 347.0898.

General Condition for Vorbrüggen Coupling Reaction. All operations were carried out under an argon protected atmosphere.

Silylation of Nucleobases. The nucleobase $(N^6$ -Benzoyl protected in case of adenine) (2 equiv) was suspended in hexamethyldisilazane (50 equiv) containing trimethylsilyl chloride (0.7 equiv) and pyridine (10 equiv). The mixture was heated at reflux overnight. After cooling, the solvent was evaporated and dried under a high vacuum.

Coupling at Ambient Condition (A). To the silylated nucleobase was added compound $17/19/20/24$ or 28 (1 equiv) dissolved in dry 1,2-dichloroethane (7 mL/mmol), and trimethylsilyl triflate or anhydrous $SnCl₄$ (2.5 equiv) was added dropwise at room temperature. The clear solution was stirred at rt.

Coupling under Microwave Condition (B). To the silylated nucleobase was added compound 17/19/20/24 or 28 (1 equiv) dissolved in dry acetonitrile (7 mL/mmol) , followed by the addition of trimethylsilyl triflate (0.2 equiv) at rt. The clear solution was irradiated to microwave (continuous power, 300 W, preheating $0 \rightarrow 150$ °C in 3 min, at 150 ± 3 °C for 5 min).

Workup Procedure. The reaction mixture was quenched with saturated aqueous NaHCO_3 and extracted with ethyl acetate (3 times). The combined organic layers were dried over anhydrous $Na₂SO₄$ and evaporated. Purification of the residue by silica-gel flash column chromatography (MeOH–CH₂Cl₂) afforded the pure coupled product as white foam.

1′-(Thymin-1-yl)-2′-O-acetyl-3′-deoxy-5′-O-benzyl-α-D-apio-Lfuranose (29). Using condition A, compound 19 (320 mg, 1.04 mmol) gave compound 29 (420 mg) in quantitative yield as a white foam: ¹H NMR (300 MHz, CDCl₃) δ ppm 1.84 (d, J = 0.9 Hz, 3H, 5-CH3), 1.98 (s, 3H, 2′-OAc), 2.67−2.85 (m, 1H, 3′-H), 3.39 (dd, J = 9.1, 7.62 Hz, 1H, 5′-H), 3.57 (dd, J = 9.2, 6.0 Hz, 1H, 5′-H), 3.89 (t, J = 8.9 Hz, 1H, 4′-H), 4.36 (t, J = 8.1 Hz, 1H, 4′-H), 4.43 (s, 2H, CH₂Ph), 5.39 (dd, J = 6.2, 2.3 Hz, 1H, 2'-H), 5.74 (d, J = 2.3 Hz, 1H, 1′-H), 6.89−7.02 (d, J = 0.9 Hz, 1H, 6-H), 7.21−7.38 (m, 5H, CH₂Ph), 8.85 (br s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm 12.6 (5-CH3), 20.6 (2′−OCOCH3), 41.0 (3′-C), 66.3 (5′-C), 71.9 (4′- C), 73.5 (CH₂Ph), 91.2 (1'-C), 111.0 (5-C), 127.7 (CH₂Ph), 127.9 (CH₂Ph), 128.5 (CH₂Ph), 135.1 (6-C), 137.6 (CH₂Ph), 150.0 (2-C), 163.7 (4- C), 169.7 (2'-OCOCH₃); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for $C_{19}H_{23}N_2O_6$ 375.1556, found 375.1556.

1'-(N⁶-Benzoyladenin-9-yl)-2'-O-acetyl-3'-deoxy-5'-O-benzyl-α- D -apio-L-furanose (30). Using condition B, compound 20 (1.0 g, 3.56 mmol) gave compound 30 (1.0 g, 60%) as a white foam: $^1\rm H$ NMR (300 MHz, CDCl3) δ ppm 2.00 (s, 3H, 2′-OAc), 3.11−3.25 (m, 1H, 3′-H), 3.49 (dd, J = 9.2, 7.2 Hz, 1H, 5′-H), 3.63 (dd, J = 9.4, 6.4 Hz, 1H, 5'-H), 4.00 (t, J = 8.5 Hz, 1H, 4'-H), 4.45 (s, 2H, PhCH₂), 4.50 (t, $J = 8.1$ Hz, 1H, 4'-H), 5.79 (dd, $J = 5.9$, 2.1 Hz, 1H, 2'-H), 6.04 (d, $J =$ 2.3 Hz, 1H, 1′-H), 7.21–7.32 (m, 5H, CH₂Ph), 7.37–7.47 (m, 2H, H_m Bz), 7.47−7.56 (m, 1H, H_p Bz), 7.90−7.97 (m, 2H, H_p Bz), 7.98 (s, 1H, 8-H), 8.72 (s, 1H, 2-H), 9.11 (s, 1H, NH); 13C NMR (75 MHz, CDCl3) δ ppm 20.6 (2′−OCOCH3), 41.0 (3′-C), 66.2 (5′-C), 72.1 (4′-C), 73.4 (CH2Ph), 77.0 (2′-C), 90.2 (1′- C), 123.6 (5-C), 127.7 $(C_o, C_p$ Bn), 127.8 $(C_o$ Bz), 128.4 $(C_m$ Bn), 128.8 $(C_m$ Bz), 132.7 $(C_p$ Bz), 133.6 (C_{ips0} Bz), 137.6 (C_{ips0} Bn), 141.3 (8-C), 149.5 (6-C), 151.2 (4-C), 152.8 (2-C), 164.6 (N⁶Bz-CO), 169.9 (2'-OCOCH₃); HRMS (ESI-TOF) m/z $[M + H]^+$ Calcd for $C_{26}H_{26}N_5O_5$ 488.1934, found 488.1937.

1'-(N⁶-Benzoyladenin-1-yl)-3'-deoxy-5'-O-benzyl-α-D-apio-L-furanose (31). Spectral data: ¹H NMR (300 MHz, CDCl₃) δ ppm 2.45− 2.61 (m, 1H, 3'-H), 3.68 (dd, J = 9.4, 6.4 Hz, 1H, 5'-H), 3.76 (dd, J = 9.5, 6.3 Hz, 1H, 5′-H), 4.22 (t, J = 8.8 Hz, 1H, 4′-H), 4.41 (t, J = 8.2 Hz, 1H, 4′-H), $4.45-4.52$ (m, 2H, CH₂Ph), 4.62 (d, $J = 5.0$ Hz, 1H, 2′-H), 6.56 (s, 1H, 1′-H), 7.19−7.28 (m, 5H, CH2Ph), 7.29−7.37 (m, 2H, H_m Bz), 7.39−7.47 (m, 1H, H_n Bz), 7.97 (s, 1H, 8-H), 8.16−8.22

(m, 2H, H_o Bz), 8.40 (s, 1H, 2-H), 12.45 (br s, 1H, NH); ¹³C NMR (75 MHz, CDCl3) δ ppm 41.2 (3′-C), 65.9 (5′-C), 72.1 (4′-C), 73.6 $(CH₂Ph)$, 77.6 (2'-C), 96.6 (1'-C), 114.7 (5-C), 127.9 (C_o Bn), 128.2 $(C_n$ Bn), 128.5 $(C_m$ Bz), 128.8 $(C_m$ Bn), 129.9 $(C_o$ Bz), 132.4 $(C_n$ Bz), 137.5 (C_{ipso} Bz), 137.8 (C_{ipso} Bn), 142.0 (8-C), 142.2 (2-C), 148.8 (6-C), 158.0 (4-C), 175.5 (Bz CO); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for $C_{24}H_{24}N_5O_4$ 446.1828, found 446.1839.

1′-(Thymin-1-yl)-3′-deoxy-β-D-apio-L-furanose (33). Spectral data for the compound mixture 33 $(\text{minor}) + 5$ a: ^1H NMR $(300 \text{ MHz},$ DMSO- d_6) δ ppm 1.77 (d, J = 0.9 Hz, 1.06H, minor 5-CH₃), 1.80 (d, J = 1.2 Hz, 2.89H, 5-CH3, major), 2.22−2.36 (m, 1H, 3′-H, major), 2.52−2.60 (m, 0.29H, 3′-H, minor) 3.40−3.52 (m, 1.43H, 5′-H, major and minor), 3.62−3.72 (m, 1.43H, 5′-H, major and minor), 3.73−3.81 (m, 1.11H, 4′-H, major), 3.81−3.87 (m, 0.31H, 4′-H, minor), 9.95− 4.02 (t, J = 7.9 Hz, 0.39H, 4′-H, minor), 4.11−4.16 (m, 0.36H, 2′-H, minor), 4.19 (td, $J = 5.1$, 2.1 Hz, 1.05H, 2'-H, major), 4.33 (t, $J = 7.9$ Hz, 1H, 4′-H, major), 4.51 (t, J = 5.1 Hz, 1.34H, 5′−OH, major and minor), 5.29 (d, J = 4.7 Hz, 0.37H, 2′−OH, minor), 5.51 (d, J = 5.0 Hz, 1.02H, 2′−OH, major), 5.61 (d, J = 2.1 Hz, 1H, 1′-H, major), 5.88 $(d, J = 3.2 \text{ Hz}, 0.36\text{H}, 1'$ -H, minor), 7.30 $(d, J = 1.2 \text{ Hz}, 0.36\text{H}, 6\text{-H},$ minor), 7.38 (d, J = 1.2 Hz, 1.02H, 6-H, major), 11.21 (s, 0.39H, NH, minor), 11.27 (s, 1.01H, major); ¹³C NMR (75 MHz, DMSO- d_6) δ ppm 12.1 (5-CH3, major), 12.2 (5-CH3, minor), 43.6 (3′-C, major), 46.1 (3′-C, minor), 57.6 (5′-C, major), 57.7 (5′-C, minor), 69.3 (2′-C, minor), 69.8 (4′-C, minor), 71.2 (4′-C, major), 74.2 (2′-C, major), 87.9 (1′-C, minor), 92.2 (1′-C, major), 106.8 (5-C, minor), 108.9 (5- C, major), 135.7 (6-C, major), 138.0 (6-C, minor), 150.3 (2-C, major), 150.4 (2-C, minor), 163.9 (4-C, major), 164.1 (4-C, minor).

1′-(Thymin-1-yl)-2′,3′-di(O-acetyl)-5′-O-benzyl-α-D-apio-L-furanose (34). Using condition A, compound 24 (100 mg, 0.27 mmol) gave compound 34 (100 mg, 85%) as a colorless glass: ¹H NMR (300 MHz, CDCl₃) δ ppm 1.95 (d, J = 1.2 Hz, 3H, 5–CH₃), 2.04 (s, 3H, Ac), 2.08 (s, 3H, Ac), 3.88 (s, 2H, 5′-H), 4.20 (d, J = 10.5 Hz, 1H, 4′-H), 4.55 (s, 2H, CH₂Ph), 4.56 (d, J = 10.5 Hz, 1H, 4'-H), 5.63 (d, J = 5.0 Hz, 1H, 2'-H), 5.96 (d, J = 5.0 Hz, 1H, 1'-H), 7.28 (d, J = 1.2 Hz, 1H, 6-H), 7.30−7.41 (m, 5H, CH₂Ph), 8.52 (s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm 12.7 (5-CH₃), 20.5 (Ac-CH₃), 21.6 (Ac- $CH₃$), 66.7 (5′-C), 73.2 (4′-C), 73.8 (CH₂Ph), 78.1 (2′-C), 86.2 (3′-C), 88.2 (1'-C), 111.4 (5-C), 127.8 (C_o Bn), 128.0 (C_o Bn), 128.5 (C_m Bn), 135.0 (6-C), 137.3 (C_{ipso} Bn), 150.2 (2-C), 163.3 (4-C), 169.1 (Ac-CO), 169.9 (Ac-CO); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for $C_{21}H_{25}N_2O_8$ 433.1605, found 431.1603.

 $1'$ -(N⁶-Benzoyladenin-9-yl)-2',3'-di(O-acetyl)-5'-O-benzyl- α -Dapio-L-furanose (35). Using condition B, compound 24 (220 mg, 0.6 mmol) gave compound 35 (130 mg, 40%) and 36 (20 mg, 6%): ¹H NMR (300 MHz, CDCl₃) δ ppm 2.04 (s, 3H, Ac), 2.07 (s, 3H, Ac), 3.94−4.04 (2d, J = 10.0 Hz, 2H, 5′-H), 4.37 (d, J = 10.5 Hz, 1H, 4′- H), 4.59 (s, 2H, CH₂Ph), 4.71 (d, J = 10.5 Hz, 1H, 4'-H), 6.13 (d, J = 4.4 Hz, 1H, 2′-H), 6.17 (d, J = 4.1 Hz, 1H, 1′-H), 7.31−7.39 (m, 5H, CH₂Ph), 7.51–7.56 (m, 2H, H_m Bz), 7.58–7.62 (m, 1H, H_n Bz), 8.00−8.05 (m, 2H, H_o Bz) 8.23 (s, 1H, 8-H), 8.82 (s, 1H, 2-H), 9.01 (s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm 20.5 (Ac- CH₃), 21.5 (Ac-CH₃), 66.3 (5′-C) 73.8 (CH₂Ph & 4′-C), 78.7 (2′-C), 86.4 $(3'-C)$, 87.9 (1'-C), 123.1 (5-C), 127.8 (C_o Bn) 127.8 (C_o Bz), 128.1 (Cp Bn), 128.5 (C_m Bn), 128.9 (C_m Bz), 132.8 (C_m Bz), 133.6 (C_{ipso} Bn), 137.3 (C_{ipso} Bz), 141.1 (8-C), 149.5 (4-C), 151.8 (6-C), 152.9 (2-C), 164.5 (N⁶COPh), 168.9 (COCH₃), 169.9 (COCH₃); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₂₈H₂₈N₅O₇ 546.1989, found 546.2000. Spectral data for compound 1'-(N⁶-benzoyladenin-9-yl)-2'-(O-trimethylsilyl)-3′-(O-acetyl)-5′-O-benzyl-α-D-apio-L-furanose (36): ¹ ¹H NMR (300 MHz, CDCl₃) δ ppm 0.14 (s, 9H, 2'-OSi(CH₃)₃) 1.89 $(s, 3H, 3'$ -Ac) 3.96 (d, J = 10.0 Hz, 1H, 5'-H) 4.05 (d, J = 9.7 Hz, 1H, 5′-H) 4.34 (d, $J = 10.5$ Hz, 1H, 4′-H) 4.49 (d, $J = 11.7$ Hz, 1H, CH_2Ph) 4.57 (d, J = 12.0 Hz, 1H, CH_2Ph) 4.71 (d, J = 10.5 Hz, 1H, 4′-H) 5.05 (d, J = 2.6 Hz, 1H, 2′-H) 6.04 (d, J = 2.6 Hz, 1H, 1′-H) 7.27−7.39 (m, 5H, CH2Ph) 7.50−7.56 (m, 2H, H^m Bz) 7.57−7.62 (m, 1H, H^p Bz) 8.00−8.06 (m, 2H, Ho Bz) 8.15 (s, 1H, 8-H) 8.81 (s, 1H, 2-H) 9.08 (s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm -0.1 (SiCH3), 21.6 (Ac-CH3), 66.0 (5′-C), 73.7 (CH2Ph), 74.4 (4′-C), 79.0 $(2'-C)$, 88.1 $(3'-C)$, 91.7 $(1'-C)$, 123.4 $(5-C)$, 127.7 $(C_n$ Bn), 127.85 (C_o Bz), 127.86 (C_o Bn), 128.4 (C_m Bn), 128.9 (C_m Bz), 132.8 (C_p Bz), 133.7 (C_{ipso} Bn), 137.6 (C_{ipso} Bz), 141.3 (8-C), 149.4 (4-C), 151.3 (6-C), 152.7 (2-C), 164.6 (N6 Bz-CO), 170.0 (Ac-CO); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₂₉H₃₄N₅O₆Si 576.2278, found 576.2291.

1′-(Thymin-1-yl)-3′-deoxy-5′-O-benzyl-α-D-apio-L-furanose (37). Acetyl protected compound 29 (400 mg, 1.07 mmol) was dissolved in 7 N ammonia in MeOH (15 mL). The mixture was stirred at room temperature until completion (for about 3−5 h) as indicated by TLC. Solvent was evaporated, and the residue was purified by flash column chromatography using 0.5−1% MeOH−CH₂Cl₂ to afford the title compound 37 (341 mg, 96%) as a white foam: 1 H NMR (300 MHz, CDCl₃) δ ppm 1.83 (d, J = 1.2 Hz, 3H, 5-CH₃), 2.27–2.49 (m, 1H, 3[']-H), 3.57 (dd, J = 9.2, 7.8 Hz, 1H, 5′-H), 3.78 (dd, J = 9.2, 6.0 Hz, 1H, 5′-H), 4.03 (dd, J = 10.5, 8.5 Hz, 1H, 4′-H), 4.29−4.38 (m, 2H, 4′-H and 2′-H), 4.45 (s, 2H, CH2Ph), 5.02 (br s, 1H, 2′−OH), 5.67 (s, 1H, 1′-H), 7.11 (d, J = 1.2 Hz, 1H, 6-H), 7.19–7.30 (m, 5H, CH₂Ph), 10.44 (br s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm 12.6 (5-CH₃), 41.3 (3' - C), 66.6 (5' - C), 72.9 (4' - C), 73.6 (CH₂Ph), 75.8 (2' -C), 94.3 (1′-C), 110.5 (5-C), 127.72 (CH₂Ph), 127.74 (CH₂Ph), 128.4 (CH₂Ph), 134.7 (6- C), 137.9 (CH₂Ph), 150.6 (2-C), 164.5 (4-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₇H₂₁N₂O₅ 333.1450, found 333.1458.

 $1'$ -(Adenin-9-yl)-3'-deoxy-5'-O-benzyl- α -D-apio-L-furanose (38). Compound 30 (1.0 g, 2.05 mmol) was dissolved in 7 N ammonia in MeOH (30 mL). The mixture was stirred at room temperature for 48 h. Solvent was evaporated, and the residue was purified by flash column chromatography using 2% MeOH–CH₂Cl₂ to afford the title compound 38 (650 mg, 75%) as a white foam [procedure to remove acetamide residue if any: suspend the product in water and then collect it by filtration]: ¹H NMR (300 MHz, DMSO- d_6) δ ppm 2.75–2.89 $(m, 1H, 3' - H), 3.53$ $(t, J = 8.8 \text{ Hz}, 1H, 5' - H), 3.73$ $(dd, J = 9.4, 5.9 \text{ Hz},$ 1H, $5'$ -H), 3.86 (t, J = 8.2 Hz, 1H, 4'-H), 4.40 (t, J = 7.8 Hz, 1H, 4'-H), 4.45−4.56 (m, 2H, Bn H), 4.63 (td, J = 5.3, 2.1 Hz, 1H, 2′-H), 5.76 (d, J = 4.7 Hz, 1H, 2′−OH), 5.90 (d, J = 2.3 Hz, 1H, 1′-H), 7.26 (br s, 2H, NH), 7.27−7.39 (m, 5H, CH2Ph), 8.15 (s, 1H, 2-H), 8.23 (s, 1H, 8-H); ¹³C NMR (75 MHz, DMSO- d_6) δ ppm 41.7 (3'-C), 66.8 (5′-C), 71.1 (4′-C), 72.3 (Bn C), 74.4 (2′-C), 91.1 (1′-C), 119.2 (5- C), 127.4 (C_o Bn), 127.5 (C_p Bn), 128.2 (C_m Bn), 138.5 (C_{ipso} Bn), 139.0 (8-C), 148.8 (4-C), 152.5 (2-C), 156.0 (6-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₇H₂₀N₅O₃ 342.1566, found 342.1565.

 $1'$ -(Thymin-1-yl)-5'-O-benzyl- α -D-apio-L-furanose (39). Following a similar procedure described for compound 37, compound 34 (100 mg, 0.23 mmol) gave compound 39 (81 mg, 86%) as a white foam: $^1\mathrm{H}$ NMR (300 MHz, CDCl₃) δ ppm 1.81 (d, J = 0.9 Hz, 3H, 5-CH₃), 3.60 $(d, J = 9.7 \text{ Hz}, 1\text{H}, 4'$ -H $)$, 3.85 $(d, J = 9.7 \text{ Hz}, 1\text{H}, 4'$ -H $)$, 3.90 $(d, J = 1)$ 1.2 Hz, 1H, 3′−OH), 4.06 (dd, J = 9.4, 1.5 Hz, 1H, 5′-H), 4.17 (d, J = 9.4 Hz, 1H, 5′-H), 4.41 (d, J = 3.5 Hz, 1H, 2′-H), 4.50−4.69 (app-q, J $= 12.0$ Hz, 2H, CH₂Ph), 5.25 (d, J = 3.5 Hz, 1H, 2′–OH), 5.72 (s, 1H, 1′-H), 7.23–7.37 (m, 5H, CH₂Ph), 7.52 (d, J = 1.2 Hz, 1H, 6-H), 10.81 (br s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm 12.4 (5- $CH₃$), 69.6 (4'-C), 73.7 (CH₂Ph), 77.0 (5'-C), 80.0 (2'-C), 80.5 (3'-C), 94.4 (1'-C), 108.6 (5-C), 127.77 (C_o Bn), 127.84 (C_n Bn), 128.4 $(C_m$ Bn), 137.55 $(C_{ipso}$ Bn), 137.62 (6-C) 151.3 (2-C), 164.8 (4-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₇H₂₁N₂O₆ 349.1400, found 349.1384.

 $1'$ -(Adenin-9-yl)-5'-O-benzyl- α -D-apio-L-furanose (40). Following a similar procedure described for compound 38, compound 35 (120 mg, 0.22 mmol) gave compound 40 (73 mg, 93%) as a white foam. The same procedure was employed to convert 36 to 40: ¹H NMR (300 MHz, DMSO- d_6) δ ppm 3.57–3.70 (2d, J = 9.7 Hz, 2H, 5′-H), 4.00 (d, J = 8.8 Hz, 1H, 4'-H), 4.11 (d, J = 9.1 Hz, 1 H4'-H), 4.39 (dd, $J = 5.3$, 2.9 Hz, 1H, 2′-H) 4.51–4.64 (2d, $J = 12.3$ Hz, 2H, CH₂Ph), 5.59 (s, 1H, 3′−OH), 5.90 (d, J = 2.9 Hz, 1H, 1′-H), 5.97 (d, J = 5.6 Hz, 1H, 2′−OH), 7.27 (s, 2H, NH), 7.28−7.43 (m, 5H, CH₂Ph) 8.15 (s, 1H, 2-H), 8.29 (s, 1H, 8-H); ¹³C NMR (75 MHz, DMSO- d_6) δ ppm 71.1 (5′-C), 72.7 (CH2Ph), 75.6 (4′-C), 79.8 (3′-C), 80.3 (2′-C), 90.7 (1'-C), 118.8 (5-C), 127.3 (C_p Bn), 127.4 (C_o Bn), 128.2 (C_m Bn), 138.5 (Cipso Bn), 139.7 (8-C), 149.0 (4-C), 152.5 (2-C), 156.0 (6C); HRMS (ESI-TOF) m/z $[M + H]^+$ Calcd for $C_{17}H_{20}N_5O_4$ 358.1515, found 358.1512.

1'-(Thymin-1-yl)-3'-deoxy-α-p-apio-L-furanose (5a).¹¹ Compound 37 (300 mg, 0.9 mmol) was dissolved in MeOH (10 mL), to this was added Pd−C (300 mg, 10% Pd, wet ∼50%). A stream [of h](#page-14-0)ydrogen gas was bubbled through the reaction mixture with vigorous stirring for about 1 h, and the mixture was then stirred under hydrogen atmosphere overnight at room temperature. The catalyst was filtered off, the filtrate was concentrated and purified by silica-gel flash column chromatography eluting with 6-8% MeOH-CH2Cl2 to afford compound $5a$ (190 mg, $86\%)$ as a white solid: ¹H NMR (300 MHz, DMSO- d_6) δ ppm 1.79 (d, J = 1.2 Hz, 3H, 5-CH₃), 2.22–2.36 (m, 1H, 3′-H), 3.46 (ddd, J = 10.8, 7.7, 5.3 Hz, 1H, 5′-H), 3.62−3.71 (m, 1H, 5'-H), 3.76 (t, $J = 8.6$ Hz, 1H, 4'-H), 4.18 (td, $J = 5.0$, 2.1 Hz, 1H, 2'-H), 4.33 (t, J = 7.8 Hz, 1H, 4′-H), 4.51 (t, J = 5.1 Hz, 1H, 5′–OH), 5.51 (d, J = 4.7 Hz, 1H, 2′–OH), 5.61 (d, J = 2.1 Hz, 1H, 1′-H), 7.38 (d, $J = 1.2$ Hz, 1H, 6-H), 11.27 (br s, 1H, NH); ¹³C NMR (75 MHz, DMSO-d6) δ ppm 12.1 (5-CH3), 43.6 (3′-C), 57.6 (5′-C), 71.1 (4′- C), 74.2 (2′-C), 92.2 (1′-C), 108.9 (5-C), 135.7 (6-C), 150.3 (2-C), 163.9 (4-C); HRMS (ESI-TOF) m/z $[M + H]^+$ Calcd for $C_{10}H_{15}N_2O_5$ 243.0981, found 243.0990.

 $1'$ -(Adenin-9-yl)-3'-deoxy- α -p-apio-L-furanose (5b). Compound 38 (450 mg, 1.32 mmol) was dissolved in 1:1 v/v mixture of MeOH–formic acid (40 mL), to this was added Pd(OH)₂-C (300 mg, 10% Pd, wet ∼50%) and stirred at 55 °C for 5−8 h. The catalyst was filtered off, and the filtrate was concentrated. The residue contained compound 5b and 41 as a mixture. The residue was dissolved in 7 N NH₃−MeOH and stirred at room temperature for 3 h. The volatiles were evaporated, and the residue purified by silica-gel flash column chromatography eluting with 10−12% MeOH−CH₂Cl₂ to afford compound 5b (265 mg, 80%) as a white solid. Spectral data for 1′- (adenin-9-yl)-3'-deoxy-5'-O-formyl- α -D-apio-L-furanose (41): ¹H NMR (300 MHz, DMSO-d₆) δ ppm 2.82−3.00 (m, 1H, 3'-H), 3.86 (t, J = 8.2 Hz, 1H, 4′-H), 4.21 (dd, J = 11.0, 7.8 Hz, 1H, 5′-H), 4.32− 4.46 (m, 2H, 4′ and 5′-H's), 4.70 (br s, 1H, 2′-H), 5.93 (d, J = 2.1 Hz, 1H, 1′-H), 7.28 (s, 2H, NH), 8.15 (s, 1H, 2-H), 8.24 (s, 1H, 5′− OCOH), 8.25 (s, 1H, 8-H); ¹³C NMR (75 MHz, DMSO- d_6) δ ppm 41.3 (3′-C), 61.3 (5′-C), 71.1 (4′-C), 74.7 (2′-C), 91.8 (1′-C), 119.7 (5-C), 139.9 (8-C), 149.4 (4-C), 153.3 (2-C), 156.5 (6-C), 162.8 (5′- OCOH); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₁H₁₄N₅O₄ 280.1046, found 280.1046. Spectral data for 1-(adenin-9-yl)-3′-deoxy- α -D-apio-L-furanose (5b): ¹H NMR (300 MHz, DMSO- d_6) δ ppm 2.53−2.66 (m, 1H, 3′-H), 3.52 (t, J = 8.8 Hz, 1H, 5′-H), 3.67−3.77 $(m, 1H, 5'$ -H), 3.86 $(t, J = 8.2 \text{ Hz}, 1H, 4'$ -H), 4.35 $(t, J = 7.8 \text{ Hz}, 1H,$ 4′-H), 4.54 (br s, 1H, 5′–OH), 4.63 (br s, 1H, 2′-H), 5.64 (d, J = 4.7) Hz, 1H, 2′−OH), 5.89 (d, J = 2.3 Hz, 1H, 1′-H), 7.25 (br s, 2H, NH₂), 8.15 (s, 1H, 2-H), 8.22 (s, 1H, 8-H); ¹³C NMR (75 MHz, DMSO- \tilde{d}_6) δ ppm 44.1 (3′-C), 57.6 (5′-C), 70.8 (4′-C), 74.4 (2′-C), 91.1 (1′-C), 119.2 (5- C), 138.9 (8-C), 148.8 (4-C), 152.5 (2-C), 156.0 (6-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₀H₁₄N₅O₃ 252.1097, found 252.1090.

 $1'$ -(Thymin-1-yl)- α -D-apio-L-furanose (6a). Following a similar procedure described for compound 5a, compound 39 (210 mg, 0.60 mmol) gave compound 6a (110 mg, 71%) as a white foam: ¹H NMR $(300 \text{ MHz}, \text{DMSO-}d_6) \delta \text{ ppm} 1.77 \text{ (d, } J = 1.2 \text{ Hz}, 3H, 5\text{-CH}_3)$, 3.54 (s, 2H, 5′-H), 3.88 (d, J = 9.1 Hz, 1H, 4′-H), 3.93 (br s, 1H, 2′-H), 3.98 (d, J = 9.1 Hz, 1 H), 4.57 (br s, 1H, 3′−OH), 5.04 (s, 1H, 5′−OH), 5.67 (d, J = 2.6 Hz, 1H, 1′-H), 5.72 (d, J = 4.7 Hz, 1H, 2′−OH), 7.62 $(d, J = 1.2 \text{ Hz}, 1H, 6-H), 11.25 \text{ (br s, 1H, NH)}.$ ¹³C NMR (75 MHz, DMSO- d_6) δ ppm 13.0 (5-CH₃), 63.0 (5'-C), 76.5 (4'-C), 80.5 (2'-C), 81.0 (3′-C), 93.0 (1′-C), 108.7 (5-C), 137.9 (6-C), 151.2 (2-C), 164.6 (4-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₀H₁₅N₂O₆ 259.0930, found 259.0927.

 $1'$ -(Adenin-9-yl)- α -D-apio-L-furanose (6b).⁴⁶ Compound 40 (20 mg, 0.056 mmol) was dissolved in 9:1 v/v mixture MeOH−formic acid (2 mL), to this was added $Pd(OH)_{2}$ -[C \(](#page-15-0)20 mg, 10% Pd, wet \sim 50%) and stirred at 55 °C for 5−8 h. The catalyst was filtered off, the filtrate was concentrated, and the residue was purified by silica-gel flash column chromatography eluting with 10−14% MeOH−CH₂Cl₂ to afford compound $6b$ (12 mg, 80%) as a white solid: ¹H NMR (300

MHz, DMSO- d_6) δ ppm 3.62 (d, J = 5.6 Hz, 2H, 5'-H), 3.98 (d, J = 9.1 Hz, 1H, 4′-H), 4.04 (d, J = 9.1 Hz, 1H, 4′-H), 4.38 (br s, 1H, 2′- H), 4.64 (t, J = 5.7 Hz, 1H, 5′−OH), 5.36 (s, 1H, 3′−OH), 5.85 (d, J = 4.7 Hz, 1H, 2′−OH), 5.90 (d, J = 2.9 Hz, 1H, 1′-H), 7.26 (s, 2H, NH), 8.15 (s, 1H, 2-H), 8.31 (s, 1H, 8-H); 13C NMR (75 MHz, DMSO-d6) δ ppm 62.0 (5′-C), 75.3 (4′-C), 80.1 (2′-C), 80.3 (3′-C), 90.6 (1′-C), 118.7 (5-C), 139.6 (8-C), 149.0 (4-C), 152.3 (2-C), 155.9 (6-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₀H₁₄N₅O₄ 268.1046, found 268.1036.

1′-(Thymin-1-yl)-3′-deoxy-5′-O-benzyl-β-D-apio-D-furanose (42). Using Vorbrüggen coupling condition A and then following procedure described for 37, compound 28 (550 mg, 1.78 mmol) gave 42 (360 mg, 60%) as a white foam: 1 H NMR (300 MHz, CDCl₃) δ ppm 1.84 $(d, J = 0.7 \text{ Hz}, 3\text{H}, 5-\text{CH}_3), 2.68 \text{ (ddt, } J = 12.7, 7.7, 6.4 \text{ Hz}, 1\text{H}, 3'-\text{H}),$ 3.51 (dd, J = 9.5, 6.6 Hz, 1H, 5′-H), 3.59 (dd, J = 9.5, 5.0 Hz, 1H, 5′- H), 4.01 (dd, J = 8.8, 7.9 Hz, 1H, 4′-H), 4.22 (dd, J = 6.2, 3.9 Hz, 1H, 2′-H), 4.32 (dd, J = 8.8, 7.8 Hz, 1H, 4′-H), 4.50 (s, 2H, PhCH₂), 5.60 $(d, J = 3.8 \text{ Hz}, 1H, 1'$ -H $), 7.24 (d, J = 1.3 \text{ Hz}, 1H, 6$ -H $), 7.26 - 7.38 (m,$ 5H, PhCH₂), 9.77 (s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm 12.5 (5-CH₃), 46.4 (3'-C), 68.6 (5'-C), 71.4 (4'-C), 73.3 (PhCH₂), 79.2 (2'-C), 94.3 (1'-C), 110.4 (5-C), 127.7, 127.9, 128.5 (PhCH₂) 135.0 (6-C) 137.8 (PhCH₂), 151.6 (2-C), 164.1 (4-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₇H₂₁N₂O₅ 333.1450, found 333.1452.

1′-(Adenin-9-yl)-3′-deoxy-5′-O-benzyl-β-D-apio-D-furanose (43). Using Vorbrüggen coupling condition B and then following procedure described for 38, compound 28 (1.55 g, 5 mmol) gave 43 (480 mg, 28%) and its α -anomer (200 mg, 11%) as a white solid: ¹H NMR (300 MHz, DMSO- d_6) δ ppm 2.60 (quind, J = 8.1, 5.0 Hz, 1H, 3'-H), 3.61 $(t, J = 8.5 \text{ Hz}, 1\text{H}, 5' \text{-H})$, 3.70 (dd, $J = 9.7, 5.0 \text{ Hz}, 1\text{H}, 5' \text{-H}$), 4.05 (t, J $= 8.8$ Hz, 1H, 4′-Hb), 4.17 (t, $J = 8.2$ Hz, 1H, 4′-Ha), 4.51 (s, 2H, PhCH₂), 4.70 (dt, J = 7.6, 5.7 Hz, 1H, 2'-H), 5.69 (d, J = 5.9 Hz, 1H, 2′−OH), 5.79 (d, J = 5.6 Hz, 1H, 1′-H), 7.26 (s, 2H, NH₂), 7.29–7.40 (m, 5H, PhCH₂), 8.13 (s, 1H, 2-H), 8.31 (s, 1H, 8-H); ¹³C NMR (75 MHz, DMSO-d₆) δ ppm 46.7 (3'-C), 69.2 (5'-C), 70.5 (4'-C), 72.2 (PhCH2), 75.1 (2′-C), 90.0 (1′-C), 119.2 (5-C), 127.44, 127.46, 128.3, 138.3 (PhCH₂), 139.8 (8-C), 149.4 (4-C), 152.6 (2-C), 156.0 (6-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₇H₂₀N₅O₃ 342.1566, found 342.1553. Spectral data for 1′-(adenin-9-yl)-3′-deoxy-5′-Obenzyl- α -D-apio-D-furanose: ¹H NMR (300 MHz, DMSO- d_6) δ ppm 2.69−2.83 (m, 1H, 3′-H), 3.55 (dd, J = 9.5, 7.2 Hz, 1H, 5′-H), 3.66 $(dd, J = 9.5, 5.1 Hz, 1H, 5' - H), 3.73 (dd, J = 8.5, 7.0 Hz, 1H, 4' - Hb),$ 4.29 (q, J = 5.6 Hz, 1H, 2'-H), 4.36 (t, J = 8.2 Hz, 1H, 4'-Ha), 4.54 (s, 2H, PhCH2), 5.53 (d, J = 5.3 Hz, 1H, 2′−OH), 6.19 (d, J = 5.3 Hz, 1H, 1′-H), 7.22 (s, 2H, NH₂), 7.26–7.43 (m, 5H, PhCH₂), 8.14 (s, 1H, 2-H), 8.16 (s, 1H, 8-H); ¹³C NMR (75 MHz, DMSO-d₆) δ ppm 45.3 (3'-C), 69.2 (4'-C), 69.4 (5'-C), 72.0 (2'-C) 72.2 (PhCH₂), 84.4 $(1′-C)$, 118.2 $(5-C)$, 127.48, 127.54, 128.3, 138.3 $(PhCH₂)$, 140.2 $(8-C)$ C), 149.6 (4-C), 152.4 (2-C), 155.8 (6-C).

1′-(Thymin-1-yl)-5′-O-benzyl-β-D-apio-D-furanose (44). Using Vorbrüggen coupling condition A and then following procedure described for 37, compound 17 (500 mg, 1.36 mmol) rendered 44 (460 mg, 97%) as a white foam: ${}^{1}H$ NMR (300 MHz, CDCl₃) δ ppm 1.89 (d, J = 1.2 Hz, 3H, 5−CH3), 3.49 (d, J = 0.9 Hz, 1H, 3′−OH), 3.52 (s, 2H, 5′-H), 4.07 (d, J = 9.7 Hz, 1H, 4′-H_b), 4.24 (dd, J = 10.0, 0.9 Hz, 1H, 4′-H_a), 4.27 (dd, J = 5.6, 3.8 Hz, 1H, 2′-H), 4.38 (d, J = 4.1 Hz, 1H, 2′−OH), 4.56 (s, 2H, PhCH2), 5.71 (d, J = 5.9 Hz, 1H, 1′- H), 7.22 (d, J = 1.2 Hz, 1H, 6-H), 7.27–7.40 (m, 5H, PhCH₂), 9.18 (s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm 12.5 (5-CH₃), 71.0 (5'-C), 73.7 (PhCH₂), 75.7 (4'-C), 76.9 (2'-C), 78.1 (3'-C), 92.4 (1'-C), 111.0 (5-C), 127.8, 128.0, 128.6, 137.4 (PhCH₂), 135.5 (6-C), 151.5 (2-C), 163.7 (4-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for $C_{17}H_{21}N_2O_6$ 349.1400, found 349.1414.

1′-(Adenin-9-yl)-5′-O-benzyl-β-D-apio-D-furanose (45). Using Vorbrüggen coupling condition B and then following procedure described for 38, compound 17 (2.7 g, 7.37 mmol) rendered 45 (1.2 g, 46%) as a white foam: $^1\text{H NMR}$ (300 MHz, DMSO- d_6) δ ppm 3.53 $(q, J = 10.0 \text{ Hz}, 2\text{H}, 5'$ -H $)$, 3.83 (d, J = 9.1 Hz, 1H, 4'-H $)$, 4.36 (d, J = 10.0 Hz, 1H, 4′-H), 4.58 (s, 2H, PhCH₂), 4.89 (t, J = 7.2 Hz, 1H, 2′-H), 5.08 (s, 1H, 3′−OH), 5.53 (d, J = 6.7 Hz, 1H, 2′−OH), 5.88 (d, J = 7.6 Hz, 1H, 1′-H), 7.19−7.29 (br.s, 2H, NH2), 7.29−7.45 (m, 5H, PhCH₂), 8.14 (s, 1H, 2-H), 8.34 (s, 1H, 8-H); ¹³C NMR (75 MHz, DMSO- d_6) δ ppm 71.3 (5'-C), 72.4 (PhCH₂), 73.8 (2'-C), 74.8 (4'-C), 77.5 (3′-C), 87.8 (1′-C), 119.4 (5-C), 127.3, 127.4, 128.2, 138.4 $(PhCH₂), 140.3 (8-C), 149.6 (4-C), 152.6 (2-C), 156.1 (6-C); HRMS$ (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₇H₂₀N₅O₄ 358.1515, found 358.1516.

1'-(Thymin-1-yl)-3'-deoxy-β-p-apio-p-furanose $(2a)$.¹¹ Following the procedure described for the synthesis of 5a, compound 42 (350 mg, 1.05 mmol) gave 2a (220 mg, 86%) as a white s[olid](#page-14-0): ¹H NMR (300 MHz, CD₃OD) δ ppm 1.89 (d, J = 1.2 Hz, 3H, 5-CH₃) 2.39– 2.55 (m, 1H, 3'-H), 3.65 (dd, J = 10.8, 6.7 Hz, 1H, 5'-H), 3.73 (dd, J = 11.0, 4.8 Hz, 1H, 5′-H), 4.02−4.10 (t, J = 8.2 Hz, 1H, 4′-H), 4.17− 4.26 (m, 2H, 2' and 4'-H's), 5.72 (d, J = 5.6 Hz, 1H, 1'-H), 7.46 (d, J = 1.2 Hz, 1H, 6-H); ¹³C NMR (75 MHz, CD₃OD) δ ppm 11.2 (5-CH₃), 48.3 (3′-C), 60.5 (5′-C), 70.3 (4′-C), 75.7 (2′-C), 92.3 (1′-C), 110.4 $(5-C)$, 137.1 $(6-C)$, 151.6 $(2-C)$, 165.2 $(4-C)$; HRMS $(ESI-TOF)$ m/z $[M + H]^+$ Calcd for $C_{10}H_{15}N_2O_5$ 243.0981, found 243.0975.

1'-(Adenin-9-yl)-3'-deoxy-β-p-apio-p-furanose (2b).⁸ Following the procedure described for the synthesis of 5b, compound 43 (600 mg, 1.76 mmol) gave 2b (390 mg, 88%) as a white so[lid](#page-14-0): ¹H NMR $(300 \text{ MHz}, \text{DMSO-}d_6)$ δ ppm 2.34–2.48 (m, 1H, 3′-H), 3.56 (dd, J = 10.7, 7.8 Hz, 1H, 5'-H), 3.68 (dd, J = 10.7, 4.5 Hz, 1H, 5'-H), 4.04 (t, J $= 8.8$ Hz, 1H, 4'-H), 4.13 (t, J = 8.2 Hz, 1H, 4'-H), 4.62 (t, J = 6.4 Hz, 1H, 2′-H), 4.79 (br.s, 1H, 5′−OH), 5.61 (br.s, 1H, 2′−OH), 5.79 (d, J = 5.6 Hz, 1H, 1′-H), 7.26 (s, 2H, NH2), 8.15 (s, 1H, 2-H), 8.31 (s, 1H, 8-H); ¹³C NMR (75 MHz, DMSO-d₆) δ ppm 49.0 (3'-C), 60.2 (5'-C), 70.3 (4′-C), 75.1 (2′-C), 90.0 (1′-C), 119.2 (5-C), 139.6 (8-C), 149.5 (4-C), 152.6 (2-C), 156.0 (6-C); HRMS (ESI-TOF) m/z [M + $[H]^+$ Calcd for $C_{10}H_{14}N_5O_3$ 252.1097, found 252.1081.

1'-(Adenin-9-yl)-β-D-apio-D-furanose (3b).⁴⁶ Following the procedure described for the synthesis of 6b, compound 45 (1.2 g, 3.37 mmol) rendered title compound 3b (800 m[g, 8](#page-15-0)9%) as a white solid: ¹H NMR (300 MHz, DMSO- d_6) δ ppm 3.46 (q, J = 11.1 Hz, 1H, S'-H), 3.76 (d, J = 9.1 Hz, 1H, 4'-H), 4.31 (d, J = 9.4 Hz, 1H, 4'-H), 4.80 (t, J = 6.4 Hz, 1H, 2′-H), 4.85 (s, 1H, 3′−OH), 4.91 (br.s, 1H, 5′− OH), 5.42 (d, J = 6.4 Hz, 1H, 2′−OH), 5.88 (d, J = 7.6 Hz, 1H, 1′-H), 7.26 (s, 2H, NH₂), 8.15 (s, 1H, 2-H), 8.33 (s, 1H, 8-H); ¹³C NMR (75 MHz, DMSO- d_6) δ ppm 62.4 (5'-C), 73.4 (2'-C), 74.5 (4'-C), 78.2 (3′-C), 87.7 (1′-C), 119.3 (5-C), 139.9 (8-C), 149.7 (4-C), 152.6 (2- C), 156.0 (6-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for $C_{10}H_{14}N_5O_4$ 268.1046, found 268.1066.
1'-(Thymin-1-yl)-2',3'-(O-thiocarbonyl)-5'-(O-benzyl)- β -p-apio-p-

furanose (46). To a solution of 44 (200 mg, 0.57 mmol) in DMF (4 \pm mL) was added thiocarbonyldiimidazole (112 mg, 0.63 mmol), and the mixture was heated to 80 °C for 90 min. The volatiles were removed under reduced pressure, and the residue was purified by silica-gel column chromatography (2% MeOH in CH_2Cl_2) to afford the title thiocarbonate 46 (200 mg, 89%) as a pale yellow solid: $^1\mathrm{H}$ NMR (300 MHz, CDCl₃) δ ppm 1.94 (d, J = 1.2 Hz, 3H, 5–CH₃), 3.89 (d, J = 11.1 Hz, 1H, 5′-H), 4.17 (d, J = 10.8 Hz, 1H, 5′-H), 4.30− 4.42 (m, 2H, 4′-H), 4.57–4.71 (m, 2H, PhCH₂), 5.47 (d, J = 0.9 Hz, 1H, 1'-H), 5.82 (d, J = 1.2 Hz, 1H, 2'-H), 7.03 (d, J = 1.2 Hz, 1H, 6-H), 7.27-7.38 (m, 5H, PhCH₂), 9.35 (s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm 12.3 (5-CH₃), 67.7 (5'-C), 73.7 (PhCH₂), 77.4 (4′-C), 88.7 (2′-C), 97.5 (1′-C), 100.2 (3′-C), 112.2 (5-C), 127.6, 128.0, 128.5, 137.1 (PhCH₂), 139.4 (6-C), 151.2 (2-C), 163.6 (4-C), 189.4 (CS). MS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₈H₁₉N₂O₆S 391.0964, found 391.0958.

¹′-(Adenin-9-yl)-2′,3′-(O-thiocarbonyl)-5′-(O-benzyl)-β-D-apio-D- furanose (47). Following the procedure described for the synthesis of 46, compound 45 (300 mg, 0.84 mmol) rendered title compound 47 (260 mg, 78%) as a pale yellow solid: ¹H NMR (300 MHz, CDCl₃) δ ppm 4.03 (d, $J = 10.8$ Hz, 1H, 5′-H), 4.33 (d, $J = 11.1$ Hz, 1H, 4′-H), 4.42 (d, $J = 11.1$ Hz, $1H$, $4'$ -H), 4.45 (d, $J = 10.5$ Hz, $1H$, $5'$ -H), 4.61 , 4.75 (d, J = 12.3 Hz, 2H, PhCH₂), 5.74 (br.s, 2H, NH₂), 6.14 (s, 1H, $2'$ -H), 6.20 (s, 1H, 1′-H), 7.29–7.40 (m, 5H, PhCH₂), 7.87 (s, 1H, 8-H) 7.95 (s, 1H, 2-H); ¹³C NMR (75 MHz, CDCl₃) δ ppm 67.0 (5'-C), 73.9 (PhCH₂), 75.0 (4′-C), 88.3 (2′-C), 90.3 (1′-C), 99.6 (3′-C), 119.9 (5-C), 128.0, 128.2, 128.6, 136.9 (PhCH2), 140.3 (8-C), 149.1

 $(4-C)$, 153.0 $(2-C)$, 155.6 $(6-C)$, 189.4 (CS) ; HRMS (ESI-TOF) m/z $[M + H]^{+}$ Calcd for $C_{18}H_{18}N_{5}O_{4}S$ 400.1079, found 400.1060.

1′-(Thymin-1-yl)-2′,3′-(dideoxydidehydro)-5′-(O-benzyl)-β-Dapio-D-furanose (48). A solution of compound 46 (180 mg, 0.46 mmol) in trimethylphosphite $(P({\rm OCH}_3)_3, 8.0 \text{ mL})$ was heated to 120 °C for 6 h. The volatile materials were removed under reduced pressure and then coevoporated 2−3 times with toluene. The residue was purified by silica-gel column chromatography (0−2% MeOH in CH_2Cl_2) to afford 48 (130 mg, 90%) as a white foam: ¹H NMR (300 MHz, CDCl₃) δ ppm 1.91 (d, J = 1.2 Hz, 3H, 5–CH₃), 4.25 (s, 2H, 5′-H), 4.58 (s, 2H, PhCH₂), 4.63–4.74 (m, 1H, 4′-H), 4.77–4.90 (m, 1H, 4′-H), 5.67–5.76 (m, 1H, 2′-H) 6.91 (q, J = 1.2 Hz, 1H, 6-H), 7.00 (m, 1H, 1′-H), 7.29–7.44 (m, 5H, PhCH₂), 8.47 (br.s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm 12.6 (5-CH₃), 65.1 (5′-CH₂), 73.2 (PhCH2), 75.6 (4′-C), 90.9 (1′-C), 111.3 (5-C), 119.9 (2′-C), 127.8, 128.1, 128.6, 137.3 (PhCH₂), 135.3 (6-C), 145.4 (3'-C), 150.5 (2-C), 163.6 (4-C); HRMS (ESI-TOF) m/z $[M + Na]$ ⁺ Calcd for $C_{18}H_{19}N_2O_6S$ 337.1164, found 337.1168.

1′-(Thymin-1-yl)-2′,3′-(dideoxydihydro)-β/α-D-apio-D/L-furanose $(1a + 4a)$. Following the procedure described for the synthesis of 25, compound 48 (120 mg, 0.38 mmol) rendered 1a and 4a as inseparable mixtures in 4:1 ratio, respectively (77 mg, 89%), as a white solid.

1′-(Thymin-1-yl)-3′-deoxy-5′-O-(tert-butyldimethylsilyl)-α-D-apio-L-furanose (49). Compound 5a (150 mg, 0.62 mmol) was dissolved in DMF (3.5 mL), to this was added imidazole (85 mg, 1.24 mmol) followed by tert-butyldimethylsilyl chloride (TBSCl, 112 mg, 0.74 mmol). The mixture was stirred at room temperature for 18 h. DMF was evaporated under reduced pressure. The residue was partitioned between EtOAc and brine. Organic layer separated, dried over sodium sulfate, solvent evaporated, and the residue purified by silica-gel flash column chromatography using 1-2% MeOH-CH2Cl2 to afford compound 49 $(210 \text{ mg}, 95\%)$ as a white solid: ¹H NMR $(300$ MHz, CDCl₃) δ ppm 0.07 (2s, 6H, Si(CH₃)₂), 0.89 (S, 9H, C(CH₃)₃), 1.94 (d, J = 0.9 Hz, 3H, 5-CH₃), 2.32–2.46 (m, 1H, 3′-H), 3.84 (d, J = 10.3, 7.3 Hz, 1H, 5′-H), 3.97 (d, J = 10.3, 5.9 Hz, 1H, 5′-H), 4.10 (t, J $= 8.5$ Hz, 1H, 4′-H), 4.35 (t, J = 7.9 Hz, 1H, 4′-H), 4.39 (t, J = 4.1 Hz, 1H, 2′-H), 4.81 (d, J = 3.2 Hz, 1H, 2′−OH), 5.74 (s, 1H, 1′-H), 7.22 (d, $J = 1.2$ Hz, 1H, 6-H), 10.19 (br s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm –5.51 (Si(CH₃)₂), –5.47 (Si(CH₃)₂), 12.7 (5-CH₃), 18.2 (C(CH₃)₃), 25.9 (C(CH₃)₃), 43.5 (3'-C), 59.8 (5'-C), 72.5 (4'-C), 76.1 (2′-C), 94.6 (1′-C), 110.5 (5-C), 134.8 (6-C), 150.7 (2-C), 164.4 (4-C); HRMS (ESI-TOF) m/z $[M + H]^+$ Calcd for $C_{16}H_{29}N_2O_5S$ i 357.1846, found 357.1852.

1′-(Adenin-9-yl)-3′-deoxy-5′-O-(tert-butyldimethylsilyl)-α-D-apio-L-furanose (50). Following a similar procedure described for compound 49, compound 5b (260 mg, 1.03 mmol) afforded compound 50 (310 mg, 82%) as a white solid: ¹H NMR (300 MHz, CDCl₃) δ ppm 0.10 (s, 3H, SiCH₃), 0.10 (s, 3H, SiCH₃), 0.91 $(s, 9H, C(CH_3), 2.65-2.78$ (m, 1H, 3′-H), 3.97 (dd, J = 6.0, 1.3 Hz, 2H, 5′-H), 4.21 (dd, J = 8.4, 7.5 Hz, 1H, 4′-H), 4.39 (dd, J = 8.5, 7.3 Hz, 1H, 4'-H), 4.81 (dt, $J = 5.7$, 2.7 Hz, 1H, 2'-H), 5.15 (d, $J = 3.2$ Hz, 1H, 2′−OH), 5.94 (br s, 2H, NH), 5.97 (d, J = 2.6 Hz, 1H, 1′-H), 7.94 (s, 1H, 8-H), 8.32 (s, 1H, 2-H); ¹³C NMR (75 MHz, CDCl₃) δ ppm -5.52 (SiCH₃), -5.50 (SiCH₃), 18.2 (C(CH₃)₃), 25.8 (C(CH₃)₃), 43.3 (3′-C), 60.0 (5′-C), 71.2 (4′-C), 77.1 (2′- C), 93.0 (1′-C), 120.3 (5-C), 138.4 (8-C), 149.0 (4-C), 152.7 (2-C), 155.5 (6-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₆H₂₈N₅O₃Si 366.1961, found 366.1941.

1′-(Thymin-1-yl)-3′-deoxy-5′-O-(tert-butyldimethylsilyl)-β-D-apio-D-furanose (51). Following a similar procedure described for compound 49, compound 2a (200 mg, 0.83 mmol) afforded compound 51 (260 mg, 88%) as a white foam: ${}^{1}H$ NMR (300 MHz, CDCl₃) δ ppm 0.05 (s, 6H, Si(CH₃)₂), 0.87 (s, 9H, C(CH₃)₃), 1.92 (d, J = 1.2 Hz, 3H, 5-CH₃), 2.51–2.66 (m, 1H, 3′-H), 3.69 (dd, J = 10.5, 6.2 Hz, 1H, 5′-H), 3.75 (dd, J = 10.3, 4.7 Hz, 1H, 5′-H), 3.94− 4.07 (m, 2H, 4′-H and 2′−OH), 4.18 (ddd, J = 7.0, 4.0, 2.8 Hz, 1H, 2′- H), 4.28 (t, J = 8.4 Hz, 1H, 4'-H), 5.61 (d, J = 4.1 Hz, 1H, 1'-H), 7.27 (d, $J = 1.2$ Hz, 1H, 6-H), 9.42 (s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm –5.40, –5.35 (SiCH₃), 12.7 (5-CH₃), 18.3 (C(CH₃)₃), 25.9 (C(CH3)3), 48.4 (3′-C), 60.8 (5′-C), 71.0 (4′-C), 78.7 (2′-C), 94.4 (1'-C), 110.7 (5-C), 134.9 (6-C), 151.7 (2-C), 164.1 (4-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₆H₂₉N₂O₅Si 357.1846, found 357.1855.

1′-(Adenin-9-yl)-3′-deoxy-5′-O-(tert-butyldimethylsilyl)-β-D-apio-D-furanose (52). Following a similar procedure described for compound 49, compound 2b (350 mg, 1.39 mmol) afforded compound 52 (415 mg, 82%) as a white solid: ¹H NMR (300 MHz, CDCl₃) δ ppm 0.03, 0.04 (s's, 2 × 3H, Si(CH₃)₃), 0.85 (s, 9H, $C(CH_3)$, 2.64−2.79 (m, 1H, 3′-H), 3.76 (dd, J = 10.4, 6.3 Hz, 1H, $5'$ -H), 3.85 (dd, J = 10.4, 4.5 Hz, 1H, $5'$ -H), 4.15 (t, J = 9.1 Hz, 1H, 4'-H), 4.30−4.40 (t, J = 8.5 Hz, 1H, 4′-H), 4.52 (dd, J = 8.6, 5.7 Hz, 1H, 2′-H), 5.69 (br.s, 1H, 2′−OH), 5.79 (d, J = 5.9 Hz, 1H, 1′-H), 5.95 (s, 2H, NH2), 7.97 (s, 1H, 8-H), 8.27 (s, 1H, 2-H); 13C NMR (75 MHz, CDCl₃) δ ppm -5.53, -5.49 (SiCH₃), 18.2 (C(CH₃)₃), 25.8 $(C(CH_3)_3)$, 47.7 (3'-C), 61.1 (5'-C), 71.1 (4'-C), 77.4 (2'-C), 92.8 (1′-C), 120.1 (5-C), 138.4 (8-C), 149.2 (4-C), 152.5 (2-C), 155.5 (6- C); HRMS (ESI-TOF) m/z $[M + H]^+$ Calcd for $C_{16}H_{28}N_5O_3Si$ 366.1961, found 366.1962.

1′-(Thymin-1-yl)-2′,3′-dideoxy-5′-O-benzyl-α-D-apio-L-furanose (53). To a solution of compound 37 (250 mg, 0.75 mmol) and DMAP (184 mg, 1.5 mmol) in acetonitrile (10 mL) was added dropwise O-ptolyl chlorothionoformate (138 μ L, 0.9 mmol) at room temperature. The mixture was stirred for additional 2 h, and then the volatile organics were evaporated under reduced pressure. The residue was suspended in EtOAc and washed with water and brine. The organic layer was dried over anhydrous sodium sulfate, and the solvent evaporated to dryness. The residue obtained was suspended in toluene (25 mL), tributyltinhydride (0.51 mL, 1.88 mmol) was added followed by at 60−70 °C was added azoisobutyronitrile (AIBN, 250 mg, 1.5 mmol) and heated to 110−120 °C for 3 h. Volatile materials were evaporated, and the residue was purified by silica-gel flash column chromatography using 0.5−2% MeOH−CH₂Cl₂ to afford compound 53 (167 mg, 70%) as a white foam: ^{1}H NMR (300 MHz, CDCl₃) δ ppm 1.86 (d, J = 1.2 Hz, 3H, 5-CH₃), 2.06 (ddd, J = 13.8, 7.9, 3.8 Hz, 1H, 2′-H), 2.16 (ddd, J = 13.8, 7.5, 6.4 Hz, 1H, 2′-H), 2.52−2.68 (m, 1H, $3'$ -H), 3.36 (dd, J = 9.1, 7.3 Hz, 1H, $5'$ -H), 3.45 (dd, J = 9.1, 5.6 Hz, 1H, $5'$ -H), 3.74 (dd, J = 8.8, 7.0 Hz, 1H, 4'-H), 4.24 (dd, J = 8.8, 7.3 Hz, 1H, 4'-H), 4.45 (s, 2H, CH₂Ph), 5.96 (dd, J = 6.4, 4.1 Hz, 1H, $1'$ -H), 7.07 (d, J = 1.2 Hz, 1H, 6-H), 7.20–7.33 (m, 5H, CH₂Ph), 8.56 (br s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm 12.7 (5-CH₃), 35.9 (2′-C), 38.1 (3′-C), 70.8 (5′-C), 72.7 (4′-C), 73.4 (CH2Ph), 87.0 $(1'-C)$, 110.4 (5-C), 127.7 (CH₂Ph), 127.9 (CH₂Ph), 128.5 (CH₂Ph), 135.0 (6-C), 137.8 (CH₂Ph), 150.1 (2-C), 163.7 (4-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₇H₂₁N₂O₄ 317.1501, found 317.1499.

1′-(Adenin-9-yl)-2′,3′-dideoxy-5′-O-benzyl-α-D-apio-L-furanose (54). Following a similar procedure described for compound 53, compound 38 (45 mg, 0.13 mmol) gave compound 54 (30 mg, 70%) as a white foam: $^1\text{H NMR}$ (300 MHz, CDCl₃) δ ppm 2.32 (ddd (dt), J = 13.9, 7.1 Hz, 1H, 2′-H), 2.68 (ddd, J = 13.6, 7.8, 2.9 Hz, 1H, 2′-H), 2.81−2.95 (m, 1H, 3′-H), 3.45−3.58 (m, 2H, 5′-H), 3.90 (dd, J = 8.8, 6.4 Hz, 1H, 4'-H), 4.34 (dd, $J = 8.6, 7.5$ Hz, 1H, 4'-H), 4.53 (s, 2H, CH₂Ph), 6.11 (br s, 2H, NH), 6.29 (dd, J = 6.9, 3.1 Hz, 1H, 1'-H), 7.27−7.39 (m, 5H, CH₂Ph), 7.90 (s, 1H, 8-H), 8.32 (s, 1H, 2- H); ¹³C NMR (75 MHz, CDCl₃) δ ppm 35.5 (2'-C), 38.2 (3'-C) 71.0 (5'-C), 72.2 (4'-C), 73.3 (CH₂Ph), 85.9 (1'-C), 120.2 (5-C), 127.6 (CH₂Ph), 127.8 (CH₂Ph), 128.4 (CH₂Ph), 137.9 (CH₂Ph), 138.5 (8-C), 149.2 $(4-C)$ 152.8 (2-C) 155.5 (6-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for $C_{17}H_{20}N_5O_2$ 326.1617, found 326.1611.

1′-(Thymin-1-yl)-2′,3′-dideoxy-5′-O-(tert-butyldimethylsilyl)-α-D-
apio-L-furanose (55).³ Following a similar procedure described for compound 53, compound 49 (190 mg, 0.53 mmol) gave compound 55 (130 mg, 72%) a[s a](#page-14-0) white foam: ${}^{1}H$ NMR (300 MHz, CDCl₃) δ ppm 0.06 (s, 6H, Si $(CH_3)_2$), 0.90 (s, 9H, C(CH₃)₃), 1.94 (d, J = 1.3 Hz, 3H, 5-CH₃), 2.06 (ddd, J = 13.8, 8.2, 4.0 Hz, 1H, 2'-H), 2.25 (dt, J $= 13.8, 6.9$ Hz, 1H, 2′-H), 2.48–2.63 (m, 1H, 3′-H), 3.58 (dd, J = 10.0, 6.8 Hz, 1H, 5′-H), 3.66 (dd, J = 10.1, 5.2 Hz, 1H, 5′-H), 3.83 (dd, J = 8.8, 6.9 Hz, 1H, 4'-H), 4.26 (dd, J = 8.7, 7.2 Hz, 1H, 4'-H), 6.04 (dd, J $= 6.6, 3.9$ Hz, 1H, 1'-H), 7.16 (q, J = 1.3 Hz, 1H, 6-H), 8.88 (br s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm −5.51 (SiCH₃), −5.48 $(SiCH₃), 12.7 (5-CH₃), 18.2 (C(CH₃), 25.8 (C(CH₃), 35.3 (2'-C),$ 40.1 (3′-C), 63.4 (5′-C), 72.2 (4′-C), 87.1 (1′-C), 110.4 (5-C), 135.1 $(6-C)$, 150.2 (2-C), 163.9 (4-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for $C_{16}H_{29}N_2O_4Si$ 341.1897, found 341.1884.

1′-(Adenin-9-yl)-2′,3′-dideoxy-5′-O-(tert-butyldimethylsilyl)-α-D-
apio-L-furanose (56).³ Following a similar procedure described for compound 53, compound 50 (300 mg, 0.82 mmol) gave compound 56 (253 mg, 88%) a[s a](#page-14-0) white foam: ¹H NMR (300 MHz, CDCl₃) δ ppm 0.07 (s, 6H, Si(CH₃)₂), 0.91 (s, 9H, C(CH₃)₃), 2.34 (ddd (dt), J $= 13.7, 7.1$ Hz, 1H, 2′-H), 2.62 (ddd, J = 13.3, 7.8, 2.9 Hz, 1H, 2′-H), 2.76 (dq, J = 13.5, 6.9 Hz, 1H, 3′-H), 3.60−3.74 (m, 2H, 5′-H), 3.92 $(dd, J = 8.8, 6.4 Hz, 1H, 4'-H), 4.31 (dd, J = 8.5, 7.3 Hz, 1H, 4'-H),$ 5.70 (br s, 2H, NH), 6.30 (dd, J = 6.7, 2.9 Hz, 1H, 1′-H), 7.93 (s, 1H, 8-H), 8.36 (s, 1H, 2-H); ¹³C NMR (75 MHz, CDCl₃) δ ppm -5.45 $(Si(CH_3))$, −5.42 $(Si(CH_3))$, 18.3 $(C(CH_3)_3)$, 25.8 $(C(CH_3)_3)$, 34.9 $(2'-C)$, 40.4 $(3'-C)$, 63.5 $(5'-C)$, 71.8 $(4'-C)$, 86.1 $(1'-C)$, 120.3 $(5-$ C), 138.6 (8-C), 149.4 (4-C), 153.0 (2-C), 155.4 (6-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₆H₂₈N₅O₂Si 350.2012, found 350.2006.

1′-(Thymin-1-yl)-2′,3′-dideoxy-5′-(tert-butyldimethylsilyl)-β-D-
apio-D-furanose (57).³ Following a similar procedure described for compound 53, compound 51 (250 mg, 0.70 mmol) gave compound 57 (215 mg, 90%) [a](#page-14-0)s a white foam: ¹H NMR (300 MHz, CDCl₃) δ ppm 0.06 (s, 6H, SiCH₃), 0.89 (s, 9H, C(CH₃)₃), 1.77 (ddd, J = 13.3, 8.9, 7.2 Hz, 1H, 2′-H), 1.94 (d, J = 1.2 Hz, 3H, 5-CH3), 2.43−2.55 (m, 1H, 2′-H), 2.55−2.72 (m, 1H, 3′-H), 3.60 (dd, J = 10.3, 5.9 Hz, 1H, 5′-H), 3.67 (dd, J = 10.3, 5.0 Hz, 1H, 5′-H), 3.94 (t, J = 7.8 Hz, 1H, 4′-H), 4.07 (t, J = 8.1 Hz, 1H, 4'-H), 6.06 (dd, J = 7.0, 6.4 Hz, 1H, 1'-H), 7.21 (q, J = 1.2 Hz, 1H, 6-H), 8.31 (br.s, 1H, NH); 13C NMR (75 MHz, CDCl₃) δ ppm -5.47, -5.44 (SiCH₃), 12.6 (5-CH₃), 18.3 $(C(CH₃)₃), 25.8 (C(CH₃)₃), 34.6 (2'-C), 40.9 (3'-C), 62.6 (5'-C),$ 71.0 (4′-C), 86.6 (1′-C), 110.9 (5-C), 134.9 (6-C), 150.3 (2-C), 163.8 (4-C); HRMS (ESI-TOF) m/z $[M + H]^+$ Calcd for $C_{16}H_{29}N_2O_4Si$ 341.1897, found 341.1891.

1′-(Adenin-9-yl)-2′,3′-dideoxy-5′-(tert-butyldimethylsilyl)-β-Dapio-p-furanose $(58)^3$ Following a similar procedure described for compound 53, compound 52 (400 mg, 1.10 mmol) gave compound 58 (310 mg, 81%) [a](#page-14-0)s a white foam: ^{1}H NMR (300 MHz, CDCl₃) δ ppm 0.05 (s, 6H, SiCH3), 0.88 (s, 9H, C(CH3)3), 2.33−2.50 (m, 1H, 2′-H), 2.57−2.81 (m, 2H, 2′ and 3′-H's), 3.71 (d, J = 5.3 Hz, 2H, 5′- H), 4.04 (t, $J = 8.2$ Hz, 1H, 4'-H), 4.14 (t, $J = 7.6$ Hz, 1H, 4'-H), 5.82 $(br.s, 2H, NH₂), 6.29$ (t, J = 5.9 Hz, 1H, 1'-H), 8.05 (s, 1H, 8-H), 8.36 (s, 1H, 2-H); ¹³C NMR (75 MHz, CDCl₃) δ ppm -5.44 (SiCH₃), 18.3 (C(CH₃)₃), 25.9 (C(CH₃)₃), 34.6 (2[']-C), 41.6 (3[']-C), 63.0 (5[']-C), 71.1 (4′-C), 85.5 (1′-C), 120.2 (5-C), 138.4 (8-C), 149.7 (4-C), 153.0 (2-C), 155.5 (6-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for $C_{16}H_{28}N_5O_2Si$ 350.2012, found 350.2009.

 $1'$ -(Thymin-1-yl)-2',3'-dideoxy- α -D-apio-L-furanose (4a).³ Following the hydrogenation procedure described for compound 5a, compound 53 (150 mg, 0.47 mmol) gave compound 4a (80 mg, 63%) as a white solid. Alternatively, compound 55 (110 [m](#page-14-0)g, 0.32 mmol) was dissolved in THF (2 mL) and TBAF (1M, 0.65 mL, 0.65 mmol) was added at room temperature. The reaction mixture was stirred for 3 h, solvents evaporated, and the residue was subjected to silica-gel flash column chromatography (4-5% MeOH–CH₂Cl₂) to afford 4 a (65 mg, 89%) as a white solid: $^1\rm H$ NMR (300 MHz, DMSO d_6) δ ppm 1.80 (d, J = 0.9 Hz, 3H, 5-CH₃), 1.96–2.13 (m, 2H, 2′-H), 2.45−2.60 (m, 1H, 3′-H), 3.33−3.48 (m, 2H, 5′-H), 3.63 (dd, J = 8.2, 6.2 Hz, 1H, 4'-H), 4.22 (dd, J = 8.2, 7.0 Hz, 1H, 4'-H), 4.76 (t, J = 5.3 Hz, 1H, 5′−OH), 5.97 (dd, J = 6.4, 4.7 Hz, 1H, 1′-H), 7.43 (d, J = 1.2 Hz, 1H, 6-H), 11.24 (s, 1H, NH); ¹³C NMR (75 MHz, CD₃OD) δ ppm 12.6 (5-CH3), 36.2 (2′-C), 41.7 (3′-C), 63.9 (5′-C), 73.3 (4′-C), 88.5 (1′-C), 111.4 (5-C), 137.9 (6-C), 152.4 (2-C), 166.7 (4-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₁₀H₁₅N₂O₄ 227.1032, found 227.1041.

 $1'$ -(Adenin-9-yl)-2',3'-dideoxy- α -p-apio-L-furanose (4b).³ Compound 56 (350 mg, 1.0 mmol) was dissolved in MeOH (15 mL) in a polypropylene vessel and NH_4F (742 mg, 20 mmol) was [ad](#page-14-0)ded at room temperature. The reaction mixture was stirred at 55 °C for 48 h; $CH₂Cl₂$ (20 mL) was added to the reaction vessel and filtered. The

filtrate was evaporated, and the residue was subjected to silica-gel flash column chromatography (10-12% MeOH-CH₂Cl₂) to afford 4b (205 mg, 87%) as a white solid: ¹H NMR (300 MHz, DMSO- d_6) δ ppm 2.21 (app-q, J = 6.7, 13.5 Hz, 1H, 2′-H), 2.54 (ddd, J = 3.5, 8.2, 12.9 Hz, 1H, 2′-H), 2.76 (sep, J = 6.4 Hz, 1H, 3′-H), 3.44 (m, 2H, 5′- H), 3.75 (dd, $J = 5.3$, 8.2 Hz, 1H, 4'-H), 4.18 (t, $J = 7.9$ Hz, 1H, 4'-H), 4.82 (t, $J = 5.0$ Hz, 1H, $5′$ -OH), 6.27 (dd, $J = 3.2$, 6.7 Hz, 1H, 1′-H), 7.24 (br s, 2H, 6-NH₂'s), 8.15 (s, 1H, 2-H), 8.26 (s, 1H, 8-H); ¹³C NMR (75 MHz, DMSO- d_6) 33.9 (2'-C), 40.4 (3'-C), 62.2 (5'-C), 70.9 (4′-C), 84.3 (1′-C), 119.2 (5-C), 139.2 (8-C), 148.9 (4-C), 152.5 (2-C), 156.0 (6-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for $C_{10}H_{14}N_5O_2$ 236.1147, found 236.1131.

1'-(Thymin-1-yl)-2',3'-dideoxy-β-p-apio-p-furanose (1a).³ Following a similar procedure described for the synthesis of compound 4b, compound 57 (200 mg, 0.59 mmol) gave compound 1a [\(1](#page-14-0)15 mg, 86%) as a white solid: ¹H NMR (300 MHz, CDCl₃) δ ppm 1.71 (t, J = 4.7 Hz, 1H, 5′−OH), 1.74−1.86 (m, 1H, 2′-H), 1.94 (d, J = 1.5 Hz, 3H, 5-CH3), 2.51−2.75 (m, 2H, 2′ and 3′-H's), 3.64−3.81 (m, 2H, 5′- H), 3.98 (dd, J = 8.8, 7.0 Hz, 1H, 4′-H), 4.07−4.16 (m, 1H, 4′-H), 6.02 (t, J = 6.6 Hz, 1H, 1′-H), 7.27−7.30 (q, J = 1.4 Hz, 1H, 6-H), 8.43 (br.s, 1H, NH); ¹³C NMR (75 MHz, CDCl₃) δ ppm 12.6 (5-CH₃), 34.7 (2′-C), 40.7 (3′-C), 63.2 (5′-C), 71.2 (4′-C), 86.9 (1′-C), 110.8 (5-C), 135.3 (6-C), 150.3 (2-C), 163.6 (4-C); HRMS (ESI-TOF) m/z $[M - H]$ [–] Calcd for C₁₀H₁₃N₂O₄ 225.0881, found 225.0875.

1'-(Adenin-9-yl)-2',3'-dideoxy-α-p-apio-p-furanose (1b).³ Following a similar procedure described for the synthesis of compound 4b, compound 58 (300 mg, 0.86 mmol) gave compound 1b [\(](#page-14-0)190 mg, 94%) as a white solid: ¹H NMR (300 MHz, DMSO- d_6) δ ppm 2.25− 2.39 (m, 1H, 2′-H), 2.52−2.67 (m, 2H, 2′ and 3′-H's), 3.48−3.65 (m, 2H, 5′-H), 3.89 (t, J = 8.2 Hz, 1H, 4′-H), 4.00 (t, J = 7.9 Hz, 1H, 4′- H), 4.82 (t, J = 5.1 Hz, 1H, 5′−OH), 6.23 (t, J = 6.7 Hz, 1H, 1′-H), 7.26 (s, 2H, NH2), 8.15 (s, 1H, 2-H), 8.32 (s, 1H, 8-H); 13C NMR (75 MHz, DMSO- d_6) δ ppm 33.7 (2'-C), 41.7 (3'-C), 61.7 (5'-C), 70.8 (4′-C), 84.3 (1′-C), 119.2 (5-C), 139.1 (8-C), 149.2 (4-C), 152.5 (2- C). 156.0 (6-C); HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for $C_{10}H_{14}N_5O_2$ 236.1147, found 236.1137.

1′-(Adenin-9-yl)-2′,3′-dideoxy-β-D-apio-D-furanose triphosphate (12). Compound 1b (25 mg, 0.106 mmol) and tributylammonium pyrophosphate 60 (117 mg, 0.212 mmol) were placed in a 50 mL and a 10 mL RB flask respectively, and dried under a high vacuum for 1 h. 2-chloro-4H-1,3,2-benzodioxaphosphinin-4-one 59 (43 mg, 0.212 mmol) was placed in a separate 10 mL flask and dried briefly (10 min) under a high vacuum. Anhydrous DMF (0.25 mL) was added to each flask under argon atmosphere. Tributylamine (dried and stored over 4A molecular sieves, 0.3 mL) was added to the flask containing tributylammonium pyrophosphate (60) with stirring. The contents of this flask were added to the flask containing 2-chloro-4H-1,3,2 benzodioxaphosphinin-4-one (59) and stirring continued for 1.5 h. The cyclic phosphitodiphosphate (61) formed was added to a flask containing compound 4b in DMF. After stirred for 1.5 h, 3% iodine solution (9:1 pyridine-water, 2.25 mL) was added dropwise and stirred for 20 min followed by the addition of water (4 mL) and stirred for additional 1.5 h. 3 M NaCl solution (0.66 mL) was added to the reaction mixture. The reaction mixture was transferred to two centrifuge tubes (∼4 mL each) and absolute ethanol (16 mL) was added to each tube, shaken well and immersed in powdered dry ice for 1 h. The tubes were centrifuged (20 °C, 3200 rpm, 20 min), and the clear solution decanted to afford crude product as white solid. The crude product was dissolved in distilled water (3.0 mL) and purified using Source-15Q ion exchange HPLC (0.5 mL injection, $0 \rightarrow 5$ min, 100% H_2O ; 5 \rightarrow 40 min, 100% H_2O to 100% 1 M triethylammonium bicarbonate buffer, linear gradient @flow rate 6 mL/min). The compound eluting at 33 min (or 0.8 M triethylammonium bicarbonate buffer) was collected and lyophilized to afford triethylammonium salt of triphosphate 12 as a white solid (17 mg, 21%) as highly hygroscopic colorless solid: ¹H NMR (300 MHz, D₂O) δ ppm 1.27 (t, J = 7.3 Hz, 24H, NCH₂CH₃), 1.33 (t, J = 7.3 Hz, 3H, NCH₂CH₃), 2.42 (ddd, J = 13.6, 8.6, 6.7 Hz, 1H, 2′-H), 2.74−2.89 (m, 1H, 2′-H), 2.89−3.12 (m, 3H, 3'-H & NCH₂CH₃), 3.19 (q, J = 7.3 Hz, 14H, NCH₂CH₃), 3.54 $(q, J = 7.1$ Hz, 2H, NCH₂CH₃), 4.05 (t, $J = 8.6$ Hz, 1H, 4[']-H), 4.14 $(app-t, J = 6.2$ Hz, 2H, 5′-H), 4.28 (t, J = 8.5 Hz, 1H, 4′-H), 6.35 (t, J = 6.7 Hz, 1H, 1′-H), 8.26 (s, 1H), 8.47 (s, 1H); 13C NMR (75 MHz, D₂O) δ ppm 7.3, 8.4, 10.7 (NCH₂CH₃), 33.6 (2'-C), 39.3 (d, $J_{p-c} = 8.1$ Hz, 3'-C), 42.4, 46.8 (NCH₂CH₃), 66.3 (d, J_{p-c} = 5.9 Hz, 5'-C), 70.8 (4′-C), 85.1 (1′-C), 150.8, 154.3; ³¹P NMR (121 MHz, D₂O) δ ppm −23.28 (br.s, 1P, β-P) −11.20, −11.04 (br. d, 2P, α & γ-P); HRMS (ESI-TOF) m/z [M $-$ H] $^{-}$ Calcd for $\rm C_{10}H_{15}N_5O_{11}P_3$ 473.9986, found 473.9987.

1′-(Adenin-9-yl)-2′,3′-dideoxy-α-D-apio-L-furanose triphosphate (13). Following the reaction protocol described for the synthesis of 12, compound 4b (25 mg, 0.106 mmol) afforded triethylammonium salt of triphosphate 13 as a white solid (45 mg, 48%): ¹H NMR (300 MHz, D₂O) δ ppm 1.21 (t, J = 7.3 Hz, 36H, HN(CH₂CH₃)₃), 2.46 (dt, J = 14.4, 7.3 Hz, 1H, 2′-H), 2.66 (ddd, J = 14.1, 8.1, 3.2 Hz, 1H, 2′-H), 3.10 (q, J = 7.3 Hz, 25H, 3′-H & HN(CH₂CH₃)₃), 3.95 (dd, J = 8.9, 6.3 Hz, 1H, 4′-H), 3.99−4.12 (m, 2H, 5′-H), 4.27 (dd, J = 8.8, 7.6 Hz, 1H, 4'-H), 6.37 (dd, J = 7.0, 3.2 Hz, 1 H), 8.16 (s, 1H, 2-H), 8.28 (s, 1H, 8-H); ¹³C NMR (75 MHz, D₂O) δ ppm 8.4 (HN(CH₂CH₃)₃), 33.9 (2′-C), 38.4 (d, J_{p-c} = 8.3 Hz, 3′-C), 46.7 (HN(CH₂CH₃)₃), 66.9 (d, J_{p-c} = 6.1 Hz, 5'-C), 71.4 (4'-C), 85.3 (1'-C), 119.1 (5-C), 140.2 (8-C) 148.6 (4-C), 152.7 (2-C), 155.7 (6-C); 31P NMR (121 MHz, D₂O) δ ppm –22.64 (dd, J = 21.1, 19.6 Hz, β P), –11.04 (d, J = 19.6 Hz, αP), -6.34 (d, J = 21.1 Hz, γP); HRMS (ESI-TOF) m/z [M – H][–] Calcd for C₁₀H₁₅N₅O₁₁P₃ 473.9986, found 473.9982.

1′-(Thymin-1-yl)-2′,3′-dideoxy-α-D-apio-D-furanose [phenyl-(benzoxy-L-alaninyl)]phosphate (9a). To a solution of 1a $(0.048 \text{ g}, 0.21)$ mmol) in anhydrous THF (4 mL) was added a solution of phosphorochloridate 64a³⁸ (0.22g, 0.64 mmol) in anhydrous THF (2 mL), followed by the dropwise addition, under an argon atmosphere, of anhydro[us](#page-15-0) NMI (0.88 mL, 1.11 mmol), and the reaction mixture was stirred at room temperature for 48 h. After this period, the solvent was removed, and the residue taken up in dichloromethane and washed with 0.5 M HCl $(2 \times 15 \text{ mL})$. The combined organics were dried over MgSO₄ filtered and evaporated. The residue was purified by preparative thin layer chromatography (2000 μ m, Aldrich) using a mixture CH₂Cl₂/MeOH 95:5 v/v as eluent to give 9a $(0.040 \text{ g},\, 35\%)$ as a pale yellow foamy solid: $^1\text{H NMR}$ $(500$ MHz, CD₃OD) δ ppm 7.47, 7.46 (d, J = 2.5 Hz, 2H, H-6), 7.37–7.32 (m, 14H, Ph and CH₂Ph), 7.23–7.18 (m, 6H, Ph), 5.99 (t, J = 6.0 Hz, 1H, H-1′), 5.98 (t, J = 6.0 Hz, 1H, H-1′), 5.17–5.15 (m, 4H, CH₂Ph), 4.17−4.05 (m, 4H,CH2OP), 4.04−4.01 (m, 2H, CHCH3), 4.00−3.87 (m, 4H, CH2O), 2.79−2.73 (m, 1H, H-3′), 2.72−2.66 (m, 1H, H-3′), 2.05−2.39 (m, 2H, H-2'a), 1.89, 1.88 (d, J = 1.5 Hz, 6H, CH₃), 1.81− 1.72 (m, 2H, H-2'b), 1.38 (d, J = 7.5 Hz, 3H, CHCH₃), 1.35 (d, J = 7.5 Hz, 3H, CHCH₃); ¹³C NMR (125 MHz, CD₃OD) δ ppm 174.9 (d, J_{CP} = 5.0 Hz, CO₂Bn), 174.7 (d, J_{CP} = 5.0 Hz, CO₂Bn), 166.44, 166.42 (CO), 152.30, 152.29 (CO), 152.21 (d, $J_{CP} = 2.8$ Hz, C_{ipso} OPh), 152.16 (d, $J_{CP} = 2.8$ Hz, C_{ipso} OPh), 137.54, 137.52 (C-6), 137.32, 137.31 (CipsoOCH2Ph), 130.82, 130.80 (Ph), 129.66, 129.64, 129.43, 129.40, 129.36, 129.31 (CH₂Ph), 126.25, 126.23 (Ph), 121.5 (d, J_{CP} = 4.6 Hz, Ph), 121.4 (d, J_{CP} = 5.3 Hz, Ph), 111.7 (C-5), 88.07, 88.03, (C-1'), 71.7, 71.5 (CH₂O), 68.4 (d, J_{CP} = 5.0 Hz, CH₂OP), 68.3 (d, J_{CP} = 5.0 Hz, CH₂OP), 68.0 (CH₂Ph), 51.8, 51.6 (CHCH₃), 40.5 (d, J_{CP} = 3.8 Hz, C-3'), 40.4 (d, $J_{CP} = 3.8$ Hz, C-3'), 35.2, 35.1 (CH₂), 20.4 (d, $J_{CP} = 7.5$ Hz, CHCH₃), 20.3 (d, $J_{CP} = 7.5$ Hz, CHCH₃), 12.5 (CH₃); ^{31}P NMR (202 MHz, CD₃OD) δ ppm 3.88, 3.33; HRMS (ESI-TOF) m/z [M + Na]⁺ Calcd for C₂₆H₃₀N₃O₈PNa 566.1668, found 566.1663; HPLC ACN/H₂O 10/90 v/v to 100/0 in 30 min, $\lambda = 280$ nm, flow 1 mL/min, $t_R = 15.42$ min.

1′-(Adenin-9-yl)-2′,3′-dideoxy-α-D-apio-D-furanose [phenyl-(benzoxy-L-alaninyl)]phosphate $(9b)$. Following the reaction protocol mentioned for the synthesis of compound 9a, 1b (0.050 g, 0.21 mmol) was reacted with phosphorochloridate 64a (0.23 g, 0.66 mmol) to give **9b** (0.030 g, 26%) as a white foamy solid: ¹H NMR (500 MHz, CD₃OD) δ ppm 8.24, 8.23 (2s, 1H, H-8), 8.22, 8.21 (2s, 1H, H-2), 7.40−7.26 (m, 16H, Ph), 7.22−7.15 (m, 4H, Ph), 6.27 (t, J = 7.0 Hz, 1H, H-1′), 6.24 (t, J = 6.5 Hz, 1H, H-1′), 5.14 (s, 4H, CH₂Ph), 4.26– 4.19 (m, 4H CH₂OP), 4.10–3.96 (m, 6H, CH₂O and CHCH₃), 2.91– 2.75 (m, 2H, H-3′), 2.69- 2.57 (m, 2H, H2'a), 2.41−2.34 (m, 2H, H2'b), 1.37 (d, J = 6.5 Hz, 3H, CH₃), 1.35 (d, J = 7.0 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CD₃OD) δ ppm 175.0 (d, J_{CP} = 4.2 Hz, $CO₂Bn$), 174.7 (d, $J_{CP} = 4.6$ Hz, $CO₂Bn$), 157.3 (C-6), 153.84, 153.83 (C-2), 152.22 (d, $J_{CP} = 2.5$ Hz, C_{ipso} OPh), 152.17 (d, $J_{CP} = 2.5$ Hz, CipsoOPh), 150.4 (C-4), 140.78, 140.74 (C-8), 137.29, 137.28 $(\dot{C}_{\text{ipso}}OCH_2Ph)$, 130.80 (d, $J_{\text{CP}} = 0.7$ Hz, Ph), 130.78 (d, $J_{\text{CP}} = 0.9$ Hz, Ph), 129.60, 129.38, 129.36, 129.35, 129.30 (CH₂Ph), 126.20 (d, $J_{\rm CP}$ = 1.3 Hz, Ph), 126.17 (d, $J_{\rm CP}$ = 1.3 Hz, Ph), 121.5 (d, $J_{\rm CP}$ = 4.6 Hz, Ph), 121.4 (d, J_{CP} = 4.6 Hz, Ph), 120.70, 120.68 (C-5), 87.00, 86.98, $(C-1')$, 71.8, 71.7 (CH_2O) , 68.22 $(d, J_{CP} = 5.5 \text{ Hz}, CH_2OP)$, 68.15 $(d,$ J_{CP} = 5.5 Hz, CH₂OP), 67.99, 67.97 (CH₂Ph), 51.8 (d, J_{CP} = 1.4 Hz, CHCH₃), 51.7 (CHCH₃), 41.10 (d, J_{CP} = 7.8 Hz, C-3'), 41.05 (d, J_{CP} $= 7.8$ Hz, C-3'), 35.2, 35.1 (CH₂), 20.4 (d, J_{CP} = 7.0 Hz, CHCH₃), 20.3 (d, J_{CP} =7.0 Hz, CHCH₃); ³¹P NMR (202 MHz, CD₃OD) δ ppm 3.80, 3.28; HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for C₂₆H₃₀N₆O₆P 553.1964, found 553.1960 and $[M + Na]^+$ Calcd for $C_{26}H_{29}N_6O_6PNa$ 575.1784, found 575.1770; HPLC ACN/H₂O 10/90 v/v to 100/0 in 30 min, $\lambda = 280$ nm, flow 1 mL/min, $t_R = 14.36$ min.

1′-(Thymin-1-yl)-2′,3′-dideoxy-α-D-apio-D-furanose [phenyl-(isopropoxy-L-alaninyl)]phosphate (10a). Following the reaction protocol mentioned for the synthesis of compound 9a, 1a (0.050 g, 0.22 mmol) was reacted with phosphorochloridate 64b (0.203g, 0.66 mmol) to give 10a (0.096 g, 88%) as a pale yellow foamy solid: ¹H NMR (500 MHz, CD₃OD) δ ppm 7.58, 7.57 (2s, 2H, H-6), 7.45 (d, J = 8.0 Hz, 2H, Ph), 7.44 (d, J = 7.5 Hz, 2H, Ph), 7.33−7.28 (m, 6H, Ph), 6.10 (t, $J = 7.0$ Hz, 1H, H-1'), 6.08 (t, $J = 7.0$ Hz, 1H, H-1'), 5.10−5.03 (m, 2H, CH(CH₃)₂), 4.33−4.21 (m, 4H, CH₂OP), 4.16− 4.10 (m, 2H, CH2O), 4.01−3.96 (m, 2H,CH2O), 3.94−3.89 (m, 2H, CHCH3), 2.95−2.86 (m, 2H, H-3′), 2.64−2.56 (m, 2H, H-2'a), 1.97 (s, 6H, CH3), 1.95−1.88 (m, 2H, H-2'b), 1.43 (d, J = 7.5 Hz, 3H, CHCH₃), 1.40 (d, J = 6.5 Hz, 3H, CHCH₃), 1.34–1.30 (m, 12H, CH(CH₃)₂); ¹³C NMR (125 MHz, CD₃OD) δ ppm 175.0 (d, J_{CP} = 5.4 Hz, CO₂iPr), 174.5 (d, $J_{CP} = 4.5$ Hz, CO₂iPr), 166.46, 166.44 (CO), 152.30 (d, $J_{CP} = 3.6$ Hz, C_{ipso} OPh), 152.28 (CO), 152.25 (d, J_{CP} = 3.6 Hz, C_{ipso}OPh), 137.6 (C-6), 130.84, 130.81, 126.25, 126.23 (Ph), 121.54 (d, J_{CP} = 4.5 Hz, Ph), 121.47 (d, J_{CP} = 5.3 Hz, Ph), 111.7 $(C-5)$, 88.15, 88.12, $(C-1')$, 71.7, 71.6 (CH_2O) , 70.19, 70.16 $(CH(CH_3)_2)$, 68.4 (d, $J_{CP} = 5.4$ Hz, CH₂OP), 68.3 (d, $J_{CP} = 5.4$ Hz, CH₂OP), 51.89, 51.88 (CHCH₃), 40.55 (d, $J_{CP} = 3.5$ Hz, C-3'), 40.49 (d, $J_{CP} = 3.6$ Hz, C-3'), 35.3, 35.2 (CH₂), 22.05, 22.03, 21.98, $(CH(CH_3)_2)$, 20.6 (d, J_{CP} = 7.2 Hz, CHCH₃), 20.4 (d, J_{CP} = 7.2 Hz, CHCH₃), 12.54, 12.52 (CH₃); ³¹P NMR (202 MHz, CD₃OD) δ ppm 3.89, 3.49; HRMS (ESI-TOF) m/z $[M + Na]$ ⁺ Calcd for $C_{22}H_{30}N_3O_8$ PNa 518.1668, found 518.1653; HPLC ACN/ H_2O 10:90 v/v to 100:0 in 30 min.; $\lambda = 280$ nm, flow 1 mL/min, $t_R =$ 13.79, 13.81 min.

1′-(Adenin-9-yl)-2′,3′-dideoxy-α-D-apio-D-furanose [phenyl-(isopropoxy-*L*-alaninyl)]phosphate (10b). Following the reaction protocol mentioned for the synthesis of compound 9a, 1b (0.050 g, 0.21 mmol) was reacted with phosphorochloridate 64b (0.201g, 0.66 mmol) to give 10 $\mathbf b$ (0.054 g, 51%) as a white foamy solid: $^1\mathrm H$ NMR (500 MHz, CD3OD) δ ppm 8.27, 8.25 (2s, 2H, H-8), 8.23, 8.22 (2s, 2H, H-2), 7.35 (d, J = 8.0 Hz, 2H, Ph), 7.34 (d, J = 7.8 Hz, 2H, Ph), 7.26−7.16 (m, 6H, Ph), 6.29 (t, J = 7.0 Hz, 1H, H-1′), 6.28 (t, J = 7.0 Hz, 1H, H-1′), 5.02–4.94 (m, 2H, $CH(CH_3)_2$), 4.33 (m, 4H, CH₂OP), 4.16–4.05 (m, 4H, CH₂O), 3.93–3.88 (m, 2H, CHCH₃), 2.96−2.89 (m, 2H, H-3′), 2.75- 2.66 (m, 2H, H2'a), 2.49−2.43 (m, 2H, H2^{'b}), 1.35 (d, J = 7.0 Hz, 3H, CH₃), 1.33 (d, J = 7.0 Hz, 3H, CH₃), 1.23–1.21 (m, 12H, CH(CH₃)₂); ¹³C NMR (125 MHz, CD₃OD) δ ppm 175.0 (d, J_{CP} = 4.5 Hz, CO₂iPr), 174.5 (d, J_{CP} = 4.5 Hz, CO₂iPr), 157.4 (C-6), 153.9 (C-2), 152.27 (d, $J_{CP} = 3.4$ Hz, C_{ipso}OPh), 152.22 (d, J_{CP} = 2.6 Hz, C_{ipso}OPh), 150.4 (C-4), 140.82, 140.79 (C-8), 130.81, 126.19, 126.16 (Ph), 121.53 (d, $J_{CP} = 5.5$ Hz, Ph), 121.45 (d, $J_{CP} = 5.5$ Hz, CH Ph), 120.73, 120.70 (C-5), 87.04, 87.04, (C-1'), 71.89, 71.83 (CH₂O), 70.2 (CH(CH₃)₃), 68.3 (d, J_{CP} = 6.4 Hz, CH₂OP), 68.2 (d, $J_{CP} = 6.4$ Hz, CH₂OP), 51.9, 51.7 (CHCH₃), 41.2 (d, $J_{CP} = 7.2$ Hz, C-3'), 41.1 (d, $J_{CP} = 7.2$ Hz, C-3'), 35.2, 35.1 (CH₂), 22.00, 21.95, 21.94, (CH(CH₃)₃), 20.6 (d, J_{CP} = 6.4 Hz, CHCH₃), 20.4 (d, J_{CP} = 6.4 Hz, CHCH₃); ³¹P NMR (202 MHz, CD₃OD) δ ppm 3.81, 3.46; HRMS (ESI-TOF) m/z [M + H]⁺ Calcd for $C_{22}H_{30}N_6O_6P$ 505.1964, found 505.1960 and $[M + Na]^+$ Calcd for

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 $C_{22}H_{29}N_6O_6$ PNa 527.1784, found 527.1794; HPLC ACN/H₂O 10/90 v/v to 100/0 in 30 min, $\lambda = 280$ nm, flow 1 mL/min, $t_R = 12.61$ min. 1′-(Thymin-1-yl)-2′,3′-dideoxy-α-D-apio-L-furanose [phenyl-(benzoxy-*L*-alaninyl)]phosphate (11a). To a solution of 4a $(0.095 \text{ g}, 0.42)$ mmol) in anhydrous THF (10 mL) was added 1.0 M solution of tertbutyl magnesium chloride in THF (0.84 mL, 0.84 mmol), and the reaction mixture was stirred under an argon atmosphere for 30 min. After this period, a solution of 64a (0.30 g, 0.84 mmol) in anhydrous THF (5 mL) was added dropwise, and the reaction mixture was stirred at room temperature for 17 h. After this period, the solvent was removed, and the residue was purified by column chromatography, gradient elution of CHCl₃/MeOH = $98/2$ to $95/5$ to give 11a (0.051) g, 22%) as a white solid: ³¹P NMR (CD₃OD, 202 MHz) δ 3.80, 3.30;
¹H NMR (CD₃OD, 500 MHz) δ 7.40–7.30 (8H, m, H-6, PhO, OCH2Ph), 7.23−7.18 (3H, m, PhO, OCH2Ph), 6.01−5.99 (0.5H, m, H-1′), $5.98-5.96$ (0.5H, m, H-1′), 5.17 , 5.16 (2H, 2s, OCH₂Ph), 4.28−4.21 (1H, m, H-4′ of one diastereoisomer), 4.14−4.00 (3H, m, 3′−CH2, CHCH3), 3.75−3.69 (1H, m, H-4′ of one diastereoisomer), 2.80−2.74 (0.5H, m, H-3′ of one diastereoisomer), 2.73−2.66 (0.5H, m, H-3′ of one diastereoisomer), 2.22−2.07 (2H, m, H-2′), 1.90 (3H, 2s, 5-CH₃), 1.38 (1.5H, d, $J = 7.2$ Hz, CHCH₃ of one diastereoisomer), 1.36 (1.5H, d, $J = 7.4$ Hz, CHCH₃ of one diastereoisomer); ¹³C NMR (CD₃OD, 125 MHz) δ 12.6 (5-CH₃), 20.38 (d, J_{C-P} = 7.2 Hz, CH₃), 20.44 (d, J_{C-P} = 7.2 Hz, CH₃), 35.61, 35.64 (C-2′), 39.73, 39.79 (C-3′), 51.7, 51.8 (CHCH3), 68.00 (OCH₂Ph), 68.49 (d, J_{C−P} = 6.0 Hz, 3′-CH₂), 68.53 (d, J_{C−P} = 5.8 Hz, 3′-CH2), 72.49, 72.54 (C-4′), 88.36, 88.38 (C-1′), 111.30, 111.33 (C-5), 121.50, 121.53, 121.57, 121.61, 126.2, 128.0, 129.33, 129.37, 129.40, 129.42, 129.65, 129.66, 130.8 (arom H), 137.3 C_{ipso} Bn), 137.74, 137.76 (C-6), 152.16, 152.19, 152.23 (C-2, C_{ipso} OPh), 166.5 (C-4), 174.8 (d, J_{C-P} = 4.6 Hz, CO), 175.0 (d, J_{C-P} = 4.6 Hz, CO); HRMS (ESI-TOF) m/z [M + Na]⁺ Calcd for C₂₆H₃₀N₃O₈PNa 566.1668, found 566.1651; HPLC = H₂O/ACN from 100/0 to 0/100 in 30 min = retention time 18.24 min; $H_2O/MeOH$ from 100/0 to 0/ 100 in 30 min = retention time 25.07 min.

1′-(Adenin-9-yl)-2′,3′-dideoxy-α-D-apio-L-furanose [phenyl-(benzoxy-*L*-alaninyl)]phosphate (11b). To a solution of 4b $(0.10 \text{ g}, 0.42)$ mmol) in anhydrous THF (10 mL) and anhydrous pyridine (2 mL) was added a solution of 64a (0.45g, 1.26 mmol) in anhydrous THF (5 mL), followed by the addition dropwise under an argon atmosphere of anhydrous NMI (0.10 mL, 1.26 mmol), and the reaction mixture was stirred at room temperature for 24 h. After this period, a solution of 64a (0.30 g, 0.84 mmol) in anhydrous THF (3 mL) and anhydrous NMI (0.07 mL, 0.84 mmol) were added, and the reaction mixture was stirred at room temperature for further 24 h. After this period, the solvent was removed, and the residue was purified by column chromatography, gradient elution of CH_2Cl_2 , then $CH_2Cl_2/MeOH =$ 98/2 then 96/4 then 90/10 to give a white solid which was triturated with diethyl ether to give 11b $(0.035 \text{ g}, 15\%)$ as a white solid: ³¹P NMR (CD₃OD, 202 MHz) δ 3.86, 3.31; ¹H NMR (CD₃OD, 500 MHz) δ 8.22, 8.21, 8.20, 8.17 (2H, 4s, H-2, H-8), 7.37−7.16 (10H, m, arom H), 6.31 (0.5H, dd, J = 7.0 Hz, 3.30 Hz, H-1′ of one diastereoisomer), 6.26 (0.5H, dd, $J = 7.00$ Hz, 3.20 Hz, H-1' of one diastereoisomer), 5.16, 5.15 (2H, 2s, CH2Ph), 4.29−4.22 (1H, m, H-4′), 4.18–4.02 (3H, m, CHCH₃, 3′-CH₂), 3.86–3.78 (1H, m, H-4′), 3.03−2.89 (1H, m, H-3′), 2.65−2.56 (1H, m, H-2′), 2.35−2.24 (1H, m, H-2'), 1.39 (1.5H, d, $J = 7.0$ Hz, CH₃ of one diastereoisomer), 1.37 (1.5H, d, J = 7.2 Hz, CH₃ of one diastereoisomer); ¹³C NMR (CD₃OD, 125 MHz) δ 20.35 (d, J_{C−P} = 6.7 Hz, CHCH₃), 20.41 (d, J_{C-P} = 6.8 Hz, CHCH₃), 35.38, 35.39 (C-2′), 40.0 (d, J_{C-P} = 2.7 Hz, C-3′), 40.1 (J_{C-P} = 2.8 Hz, C-3′), 51.7, 51.8 (CHCH₃), 67.95, 67.97 (CH₂Ph), 68.6 (d, J_{C−P} = 5.7 Hz, 3′-CH₂), 68.8 (d, J_{C−P} = 5.8 Hz, 3′-CH2), 72.13, 72.15 (C-4′), 87.0 (C-1′), 120.7, 121.48, 121.52, 121.57, 121.61, 126.18, 126.21, 129.31, 129.35, 129.37, 129.6, 129.7, 130.8 (arom H), 137.3 (C_{ijso} Bn), 140.8 (C-2), 152.19 (d, J_{C-P} = 5.5 Hz, C_{ipso} OPh), 152.25 (d, J_{C−P} = 4.7 Hz, C_{ipso} OPh), 153.7, 153.8 (C-8), 157.3 (C-8), 174.8 (d, J_{C-P} = 4.7 Hz, CO), 175.0 (d, J_{C-P} = 4.5 Hz, CO); HRMS (ESI-TOF) m/z [M + Na]⁺ Calcd for C₂₆H₂₉N₆O₆PNa 575.1784, found 575.1778; HPLC = H_2O/ACN from 100/0 to 0/100 in 30 min = retention time 17.05 min.

■ ASSOCIATED CONTENT

6 Supporting Information

Biological assay procedure, copies of $^{1}H, {^{13}C}, {^{31}P}$ and 2-D NMR spectra of relevant compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

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